

# TNF- $\alpha$ and plasma albumin as biomarkers of disease activity in systemic lupus erythematosus

Helena Idborg,<sup>1</sup> Susanna Eketjäll,<sup>2,3</sup> Susanne Pettersson,<sup>4,5</sup> Johanna T Gustafsson,<sup>1</sup> Agneta Zickert,<sup>1</sup> Marika Kvarnström,<sup>1</sup> Vilija Oke,<sup>1</sup> Per-Johan Jakobsson,<sup>1</sup> Iva Gunnarsson,<sup>1</sup> Elisabet Svenungsson<sup>1</sup>

**To cite:** Idborg H, Eketjäll S, Pettersson S, *et al.* TNF- $\alpha$  and plasma albumin as biomarkers of disease activity in systemic lupus erythematosus. *Lupus Science & Medicine* 2018;5:e000260. doi:10.1136/lupus-2018-000260

► Additional material is published online only. To view please visit the journal online (<http://dx.doi.org/10.1136/lupus-2018-000260>)

HI and SE contributed equally.

Received 14 January 2018  
Revised 20 April 2018  
Accepted 4 May 2018



<sup>1</sup>Rheumatology Unit, Department of Medicine Solna, Karolinska Institutet, Karolinska University Hospital, Stockholm, Sweden

<sup>2</sup>Cardiovascular and Metabolic Diseases, Innovative Medicines and Early Development Biotech Unit, AstraZeneca, Integrated Cardio Metabolic Centre (ICMC), Karolinska Institutet, Huddinge, Sweden

<sup>3</sup>Science for Life Laboratory, Department of Clinical Neuroscience, Karolinska Institutet, Solna, Sweden

<sup>4</sup>Theme Inflammation and Infection, Karolinska University Hospital, Stockholm, Sweden

<sup>5</sup>Department of Neurobiology, Care Sciences and Society, Karolinska Institutet, Stockholm, Sweden

## Correspondence to

Professor Elisabet Svenungsson; [Elisabet.Svenungsson@ki.se](mailto:Elisabet.Svenungsson@ki.se)

## ABSTRACT

**Objectives** Composite criteria/indices are presently used to diagnose and monitor patients with systemic lupus erythematosus (SLE). Biomarkers for these purposes would be helpful in clinical practice. We therefore evaluated a large panel of cytokines and basic laboratory tests and investigated their performance as discriminators versus controls and as biomarkers of disease activity (DA).

**Methods** We examined 437 patients with SLE, fulfilling American College of Rheumatology-82 criteria, and 322 matched controls. DA was assessed according to both SLE DA Index 2000 (SLEDAI-2K) and SLE Activity Measure (SLAM). British Isles Lupus Activity Group (BILAG) was used to assess renal DA. Additionally, 132 patients self-assessed their Global Disease Activity (PtGDA). Mesoscale Discovery 30-plex cytokine assay and routine blood chemistry was performed on fasting EDTA-plasma.

**Results** Of 26 tested biomarkers, we identified TNF- $\alpha$  as the superior discriminator between patients with SLE and controls (median=4.5 pg/mL, IQR=3.1–6.2 vs median=2.3 pg/mL, IQR=2.0–2.8). The strongest correlations to SLEDAI-2K and SLAM were obtained with TNF- $\alpha$  (Spearman rho ( $\rho$ )=0.32 and  $\rho$ =0.34, respectively), partly driven by the nephritis subgroup, and with p-albumin ( $\rho$ =–0.33 and  $\rho$ =–0.31, respectively). P-albumin was decreased and TNF- $\alpha$  was increased in patients with kidney involvement (renal BILAG A/B vs C/D/E,  $p=4\times 10^{-16}$  and  $p=6\times 10^{-9}$  respectively). IP-10 was increased in patients with joint involvement (SLAM item 24 $\geq 2$  vs  $\leq 1$ ,  $p=0.0005$ ) but did not differ when comparing patients with active/inactive kidney involvement. The most powerful correlations to PtGDA was observed with p-albumin ( $\rho$ =–0.42), IL-6 ( $\rho$ =0.30) and TNF- $\alpha$  ( $\rho$ =0.29).

**Conclusion** TNF- $\alpha$  and p-albumin both performed well as discriminators between patients with SLE and controls and as proxies for DA according to both rheumatologists' and patients' assessments. In particular, renal DA was well reflected by TNF- $\alpha$ . We propose that the TNF- $\alpha$  and p-albumin merit further investigations as clinically useful biomarkers in SLE. We also observed that the pattern of activated cytokines varies with organ involvement.

## INTRODUCTION

Systemic lupus erythematosus (SLE) is an autoimmune disease that may involve many organ systems. The heterogeneity of disease manifestations and the lack of biomarkers to diagnose and monitor disease activity (DA)

## Key messages

- TNF- $\alpha$  outperformed a large set of cytokines and conventional biomarkers as discriminator between patients with SLE and controls.
- TNF- $\alpha$  and p-albumin correlated to three different measures of systemic lupus erythematosus (SLE) disease activity (DA), one of which is patient reported.
- P-albumin, an available routine measurement, merits appreciation as a biomarker of SLE DA.
- Cytokine profile varies with organ involvement.

in SLE pose difficulties in daily clinical practice and are great obstacles for the design of clinical trials. Over the years, several different indices which measure SLE-related DA have been developed and validated,<sup>1</sup> including SLE Disease Activity Index (SLEDAI),<sup>2</sup> slightly modified 2000 (SLEDAI-2K),<sup>3</sup> Systemic Lupus Activity Measure (SLAM),<sup>4</sup> European Consensus Lupus Activity Measurement (ECLAM)<sup>5</sup> and British Isles Lupus Assessment Group (BILAG) index.<sup>6</sup> These indices are time and resource consuming and rely on doctors' subjective assessments of DA and many also incorporate laboratory results. Some grade the extent of organ involvement (SLAM, BILAG) or assess the change versus previous visits (BILAG, ECLAM). SLEDAI-2K is insensitive to change since it relies on the qualitative presence/absence of clinical symptoms or laboratory results. Patients' perspectives may add a complementary view of DA, for example, as included in Systemic Lupus Activity Questionnaire (SLAQ),<sup>7</sup> but are not included in the commonly used indices.

The available indices of DA incorporate various laboratory measures but none of them use more recent measures of systemic inflammations such as the cytokines/chemokines known to be involved in SLE.<sup>8</sup> Previous studies have investigated the performance of cytokines/chemokines as biomarkers of DA<sup>9</sup>

and reported both positive associations<sup>10</sup> as well as lack of association.<sup>11 12</sup> The majority of these studies were performed on a limited number of patients and smaller sets of biomarkers, and the reported findings have usually not been further validated or applied in clinical practice.

Thus, we simultaneously evaluated the performance of a large set of cytokines together with commonly used routine laboratory tests in a large and well-characterised cohort of patients with SLE and matched population controls. Our overall aim was to accommodate an unmet clinical need of new, simple and more sensitive biomarkers to diagnose SLE and monitor DA. We specifically addressed the following three situations: (1) discrimination of patients with SLE from controls, (2) identification of biomarkers of DA which perform well in comparison to SLAM, SLEDAI-2K and Patients' assessment of Global Disease Activity (PtGDA) and (3) biomarkers that can be used to evaluate DA in specific organ systems.

## MATERIAL AND METHODS

### Patients and controls

We included consecutive patients with SLE from the Rheumatology clinics at Karolinska University Hospital and Danderyd's Hospital in Stockholm, during the inclusion period January 2004 and September 2013. All patients were >18 years old and fulfilled the 1982 revised American College of Rheumatology classification criteria<sup>13</sup> for SLE (n=437). This is a cross-sectional study and the majority of the inclusion visits replaced a regular outpatient visit and in a few cases, patients were included while in hospital for a more severe flare. We identified and individually matched controls in the national population registry for age ( $\pm 1$  year), sex/gender and residential area to the first 322 patients with SLE. A diagnosis of SLE was the only exclusion criterion among controls. We allowed other diseases, for example, rheumatoid arthritis, in order to obtain representative population samples without SLE. Controls were investigated according to the same structured protocol as the patients with SLE including interview, medical investigation, medical file review and questionnaires. Individuals with serious infections at time of blood withdrawal were excluded. Blood samples were drawn after overnight fasting. All laboratory examinations were performed on patients and controls, either on fresh samples or after storage in  $-70^{\circ}\text{C}$  (cytokines). All participants gave informed written consent to participate.

Participants underwent a structured interview and a physical examination by a rheumatologist. History and characteristics of SLE and associated manifestations, current and prior medications were obtained through interviews and medical records. SLE DA was determined with SLAM<sup>4</sup> and SLEDAI-2K.<sup>2</sup> A subgroup of patients (n=132) also answered a question of patient numeric rating scale for global disease activity (PtGDA), scale 0–10, from the questionnaire SLAQ.<sup>7 14</sup> We specifically evaluated renal involvement with British Isles Lupus Activity Group (BILAG).<sup>6</sup> Active kidney involvement was

defined by a renal BILAG score of A or B, and inactive/absent kidney disease was defined as a renal BILAG score of C, D or E. We used SLAM to measure joint and skin/mucosa involvement since SLAM, as opposed to SLEDAI, is a graded measure, which we believe captures active joint and skin involvement better than SLEDAI. Active joint involvement was defined as SLAM item 24>1, that is, at least one objective arthritis. Active skin/mucosa manifestations were considered present if the sum of SLAM items 4 and 6 was >0.

### Basic laboratory measurements

Complement factors C3 and C4 were analysed in EDTA-plasma on a Modular analyzer (Roche). High-sensitivity creative protein (CRP) and plasma (p)-albumin were measured in heparin-plasma with BN ProSpec System (Dade Behring, Deerfield, Illinois, USA). The erythrocyte sedimentation rate (ESR) was determined in citrate-plasma by the Westergren method. Anti-dsDNA levels were measured by multiplexed bead technology (Luminex) using BioPlex 2200 system (Bio-Rad, Hercules, California, USA). Laboratory tests in clinical routine, for example, urinary (u)-albumin/creatinine ratio, were performed at the SWEDAC (<http://www.swedac.se>) accredited Clinical Chemistry Laboratories at the Karolinska University Hospital.

### Mesoscale Discovery (MSD) multiplex analysis of cytokines

EDTA-plasma samples were analysed using the MSD V-PLEX Human Cytokine 30-plex kit (K15054D; Mesoscale Discovery, Gaithersburg, Maryland, USA) according to manufacturer's instruction. In short, the plasma samples were thawed at room temperature and just before analysis diluted, twice for proinflammatory cytokine and four times for chemokine analysis, in sample diluents. The diluted samples were incubated on the MSD plates for 2 hours at room temperature while shaking. Plates were washed and incubated in additional 2 hours with detection antibodies. After washing, 2 $\times$  Read buffer T was added to each well and the plates were analysed in a Sector Imager 6000. Lower limit of quantification (LLOQ) was determined for each assay plate based on the lowest standard point with a coefficient of variation <25% and an accuracy (back-calculated concentrations) of 80%–120%.

### Statistics

Patients and controls were compared using Mann Whitney U test or  $\chi^2$  test depending on distribution. To determine correlations between SLAM/SLEDAI-2K/PtGDA and investigated variables we calculated Spearman's rank correlation coefficient ( $\rho$ ). Figure 3 is based on log-transformed data of TNF- $\alpha$  to attain normal distribution. Calculations were performed using JMP software (SAS, Cary, North Carolina, USA), Excel (Microsoft Office 2010) or Prism (GraphPad Software, V.4). A two-sided  $p < 0.05$  was considered statistically significant. Bonferroni correction for multiple comparisons was applied when indicated.

**Table 1** Cohort characteristics

	Control* (n=322)	SLE* (n=437)
Demographic data		
Age (years)	48.2 (35.4–58.6)	47.2 (34.3–58.0)
Gender (Female)	92%	92%
BMI	24.4, n=253 (22.1–27.7)	24.0, n=424 (21.4–27.2)
Clinical data		
Disease duration (years)	NR	10.6 (2.8–20.9)
SLE criteria†	NR	6 (5–7)
SLAM	NR	6 (4–10)
SLEDAI-2K	NR	4 (0–7)
SLICC <sup>48</sup>	NR	1 (0–2)
BILAG Renal	NR	A/B: n=42 C/D/E: n=363
PtGDA (n=132)	NR	3 (1–7)
Nephritis ever‡	NR	41%
Other chronic disease§	Fibromyalgia: n=3 Psoriasis: n=3 Psoriatic arthritis: n=2	Fibromyalgia: n=3 Psoriasis: n=8 Rheumatoid arthritis: n=4
Medication¶		
Methotrexate	NR	5%
Hydroxychloroquine**	NR	38%
Azathioprine	NR	17%
Mycophenolate mofetil	NR	12%
Prednisolone††	NR	39% with pred. dose=0 mg 25% with pred. dose>7.5 mg

\*Median (25% quantile–75% quantile).

†American College of Rheumatology (ACR) classification criteria.

‡Nephritis according to ACR criteria.

§Chronic disease according to participants self-reported data.

¶Per cent of patients on medication. No patients were on anti-TNF therapy.

\*\*The low usage of hydroxychloroquine in this cohort is due to that the inclusion of patients started 2004, that is, at a time when the beneficial effects of hydroxychloroquine were not established.

††Low disease activity (LLDAS) dose of prednisolone set to >7.5 mg as suggested by Franklyn *et al.*<sup>49</sup>

BILAG, British Isles Lupus Assessment Group, Renal index; BMI, body mass index; NR, not reported; PtGDA, Patients Global Disease Activity; SLAM, SLE Activity Measure; SLE, systemic lupus erythematosus; SLEDAI-2K, SLE Disease Activity Index; SLICC, Systemic Lupus International Collaborating Clinics.

## RESULTS

### Patients with SLE versus controls

We studied 437 patients with SLE and 322 population controls and the demographics and basic clinical characteristics of patients and controls are presented in [table 1](#). We performed a multiplex screening of 30 cytokines and the cytokines that were detected above LLOQ in more than 25% of the patient with SLE or control samples were included (n=20) for further analysis (online supplementary table S1). Nineteen of 20 cytokines were significantly increased (Bonferroni corrected  $p < 0.05$ ) in SLE compared with controls ([table 2](#), [figure 1](#)). In addition, six standard clinical laboratory measurements (ESR, C4, C3, p-albumin, hsCRP and anti-dsDNA) were studied comparing SLE and controls ([table 2](#), [figure 1](#)). TNF- $\alpha$  showed the lowest p value comparing SLE and controls ([figure 1](#)) and when excluding

patients ever diagnosed with nephritis (41%), TNF- $\alpha$  still showed the lowest p value ( $p = 3.4 \times 10^{-44}$ ). A receiver operating characteristic curve was obtained with an area under the curve of 0.86 (95% CI 0.83 to 0.89) for discrimination of patients with SLE versus controls ([figure 2](#)).

### Correlation to disease activity

The correlation between potential biomarkers (cytokines and standard laboratory measurements) and DA indices (SLAM, SLEDAI-2K and PtGDA) are reported in [table 2](#). P-albumin and TNF- $\alpha$  correlated to all three measurements of DA by  $\rho \geq |0.3|$  and to each other with  $\rho = 0.40$ . The relationship of TNF- $\alpha$  and p-albumin to quartiles of SLEDAI-2K, SLAM and PtGDA respectively are shown in [figure 3](#) (TNF- $\alpha$ ) and online supplementary figure S1 (p-albumin). Slightly weaker correlations to all three DA

**Table 2** Investigated potential biomarkers and their correlations to disease activity

	Concentration*		P value†	Spearman's correlation‡		
	Controls (n=322)	SLE (n=437)	Control vs SLE	SLAM <sup>4</sup> (n=437)	SLEDAI-2K <sup>2</sup> (n=437)	PtGDA§ (n=132)
<b>Cytokines¶</b>						
INF- $\gamma$	6.1 (4.6–9.9)	11.7 (7.0–20.2)	<0.001	0.08 (p=0.08)	0.14 (p=0.005)	0.07 (p=0.4)
IL-8	3.0 (2.2–4.0)	4.9 (3.1–8.7)	<0.001	0.16 (p<0.01)	0.16 (p<0.01)	0.23 (p<0.01)
IL-15	2.1 (1.8–2.4)	2.9 (2.3–4.1)	<0.001	<b>0.28</b> (p<0.01)	<b>0.28</b> (p<0.01)	0.19 (p=0.03)
Eotaxin	88.4 (70.1–113.8)	133.5 (97.6–186.0)	<0.001	0.07 (p=0.14)	0.04 (p=0.4)	0.14 (p=0.1)
MCP-1	69.0 (55.0–85.0)	109.5 (83.0–152.5)	<0.001	0.23 (p<0.01)	0.23 (p<0.01)	<b>0.28</b> (p<0.01)
MDC	939.0 (764.3–1109)	844.0 (640.3–1120)	0.002	–0.10 (p=0.04)	–0.17 (p<0.01)	–0.16 (p=0.06)
MIP-1 $\beta$	43.8 (33.4–56.4)	72.7 (50.8–108.1)	<0.001	0.18 (p<0.01)	0.20 (p<0.01)	0.20 (p=0.02)
IL-10	0.3 (0.2–0.4)	0.8 (0.5–1.5)	<0.001	0.20 (p<0.01)	<b>0.27</b> (p<0.01)	0.18 (p=0.04)
IL-6	0.6 (0.5–0.9)	1.2 (0.7–2.2)	<0.001	0.23 (p<0.01)	0.24 (p<0.01)	<b>0.30</b> (p<0.01))
<b>TNF-<math>\alpha</math></b>	2.3 (2.0–2.8)	4.5 (3.1–6.2)	<0.001	<b>0.34</b> (p<0.01)	<b>0.32</b> (p<0.01)	<b>0.29</b> (p<0.01)
IL-12/IL-23p40	131.0 (99.5–179.0)	181.0 (123.0–286.0)	<0.001	0.07 (p=0.12)	0.08 (p=0.11)	–0.05 (p=0.6)
IL-16	182.0 (147.0–225.0)	214.0 (152.0–287.0)	<0.001	0.14 (p<0.01)	0.19 (p<0.01)	0.13 (p=0.2)
IL-1 $\alpha$	6.4 (3.4–11.7)	6.3 (3.2–12.8)	0.6	0.05 (p=0.4)	0.19 (p<0.01)	–0.06 (p=0.5)
IL-7	3.7 (2.5–5.5)	5.0 (3.2–8.7)	<0.001	0.11 (p=0.03)	0.16 (p<0.01)	0.10 (p=0.2)
VEGF	56.7 (40.9–81.6)	77.1 (47.7–123.5)	<0.001	0.11 (p=0.03)	0.23 (p<0.01)	0.16 (p=0.07)
Eotaxin-3	20.8 (14.9–25.7)	24.4 (19.3–35.1)	0.0008	0.02 (p=0.8)	0.08 (p=0.3)	–0.01 (p=0.9)
IP-10	352.0 (258.8–479.0)	725.0 (446.0–1309)	<0.001	0.19 (p<0.01)	0.23 (p<0.01)	<b>0.27</b> (p=0.002)
MCP-4	56.0 (41.0–80.8)	78.0 (53.8–124.0)	<0.001	–0.01 (p=0.8)	–0.03 (p=0.6)	0.04 (p=0.6)
MIP-1 $\alpha$	12.0 (8.8–17.6)	21.5 (17.3–28.6)	<0.0001	<b>0.27</b> (p<0.01)	<b>0.26</b> (p<0.01)	0.10 (p=0.3)
TARC	54.4 (37.4–84.8)	85.3 (51.9–150.3)	<0.001	0.03 (p=0.5)	0.04 (p=0.4)	0.04 (p=0.6)
<b>Standard clinical laboratory measurements</b>						
ESR	8.5 (5–13)	19 (11–34)	<0.0001	<b>0.48</b> (p<0.01)	<b>0.28</b> (p<0.01)	0.18 (p=0.04)
C4	0.21 (0.17–0.25)	0.15 (0.1–0.2)	<0.0001	–0.12 (p=0.01)	<b>–0.36</b> (p<0.01)	–0.10 (p=0.3)
C3	1.04 (0.9–1.2)	0.88 (0.7–1.0)	<0.0001	–0.10 (p<0.05)	<b>–0.31</b> (p<0.01)	–0.02 (p=0.8)

Continued

Table 2 Continued

	Concentration*		P value†	Spearman's correlation‡		
	Controls (n=322)	SLE (n=437)		SLAM <sup>4</sup> (n=437)	SLEDAI-2K <sup>2</sup> (n=437)	PtGDA§ (n=132)
P-albumin	42 (41–44)	39 (36–41)	<0.0001	<b>-0.31</b> (p<0.01)	<b>-0.33</b> (p<0.01)	<b>-0.42</b> (p<0.01)
hsCRP	0.93 (0.4–2.1)	1.7 (0.7–5.3)	<0.0001	0.21 (p<0.01)	0.16 (p<0.01)	<b>0.25</b> (p<0.01)
anti-dsDNA	4 (4.0–4.0)	5 (4.0–19.5)	<0.0001	0.21 (p<0.01)	<b>0.47</b> (p<0.01)	0.19 (P=0.03)
U-albumin/ creatinine	0.45 (0.32–0.76)	1.11 (0.52–5.48)	<0.0001	0.22 (p<0.001)	<b>0.30</b> (p<0.001)	0.12 (p=0.2)

Concentrations of investigated potential biomarkers and u-albumin/creatinine in patients with SLE and controls, their p values comparing SLE and controls and their correlations to three different measurements of disease activity are reported. Biomarkers with correlations ( $p \geq 0.25$ ) to all three disease activity measurements are highlighted in bold. Italic numbers indicates biomarkers which are included in the respective indices.

\*Median (25% quantile–75% quantile). Concentrations of cytokines are reported in pg/mL. Standard clinical laboratory measurements reported as ESR in mm/hour, Complement factor C4 in g/L, Complement factor C3 in g/L, plasma albumin (p-albumin) in g/L, high-sensitivity (hs) CRP in mg/L, anti-dsDNA units and urinary (u) albumin/creatinine ratio in mg/mmol.

†Mann Whitney U test. Not compensated for multiple testing (variables: n=26).

‡Spearman's correlation rho (p value). Spearman's  $\rho > 0.25$  are highlighted in bold. Spearman's  $\rho = [0.20–0.39]$  is considered as weak correlation and  $\rho = 0.40–0.59$  as moderate correlation.

§PtGDA, Patients Global Disease Activity as determined by SLAQ.<sup>7</sup>

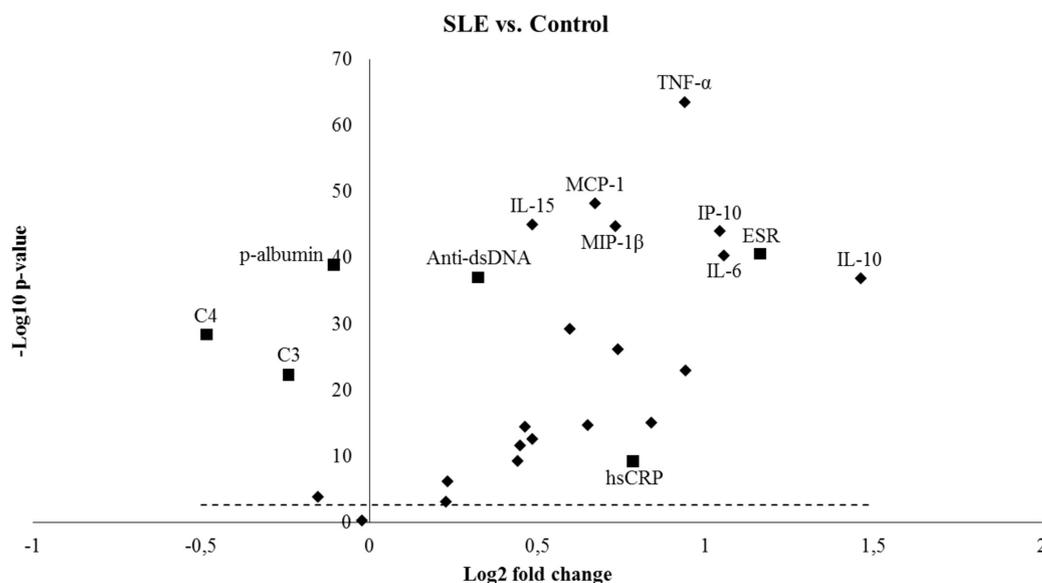
¶The numbers of missing values are reported in online supplementary table S1 (all cytokines except IL-10, IL-6, Eotaxin-3 and MIP-1 $\alpha$  were detected in more than 90% of cases). Data below LLOQ were treated as missing values.

ESR, erythrocyte sedimentation rate; LLOQ, lower limit of quantification; SLAM, SLE Activity Measure; SLAQ, Systemic Lupus Activity Questionnaire; SLE, systemic lupus erythematosus; SLEDAI-2K, SLE Disease Activity Index; VEGF, Vascular endothelial growth factor.

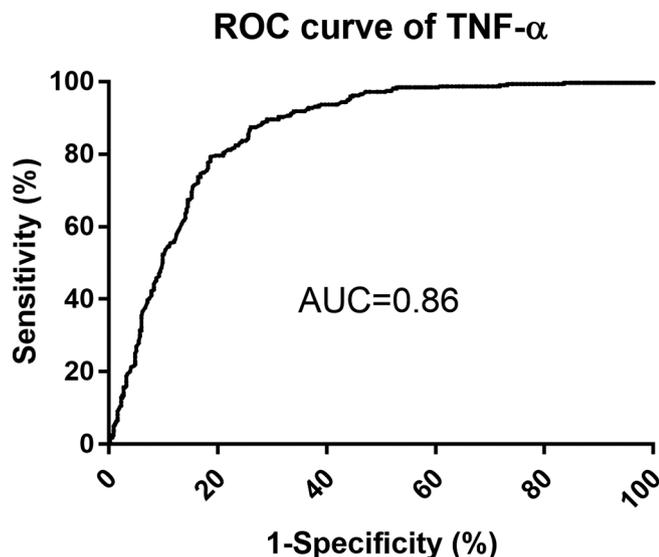
measurements were observed for IL-6 and MCP-1 ( $p \geq 0.20$ ). In addition, ESR, IL-15 and MIP-1 $\alpha$  correlated with both SLAM and SLEDAI-2K ( $p \geq 0.25$ ) but not with PtGDA. Overall, the strongest observed correlations were between p-albumin and PtGDA ( $p = -0.42$ ,  $p < 0.0001$ ) (table 2).

For SLAM, the strongest observed correlation was with ESR ( $p = 0.48$ ,  $p < 0.0001$ ), and this was expected since ESR is part of SLAM. The second strongest correlations

to SLAM ( $p = 0.31$ ) were observed for TNF- $\alpha$ , p-albumin, IL-15 and MIP-1 $\alpha$ . When we excluded all patients ever diagnosed with nephritis (online supplementary table S2), the correlation for SLAM vs. TNF- $\alpha$ , p-albumin, IL-15 and MIP-1 $\alpha$  were slightly lower ( $p = 0.21$ ). Two items included in SLEDAI-2K, anti-dsDNA ( $p = 0.47$ ) and C4 ( $p = -0.36$ ), correlated, as expected, to the total SLEDAI-2K score. The second strongest correlations to SLEDAI-2K



**Figure 1** Cytokines with the lowest p values (Mann Whitney U test) comparing SLE to controls are noted. In addition, the six investigated standard clinical laboratory measurements are shown as squares. P values above the dotted line are significant ( $p < 0.05$ ) after correction for multiple testing (n=26). SLE, systemic lupus erythematosus.



**Figure 2** ROC curve analysis of TNF- $\alpha$  was used to assess the discrimination between patients with SLE and controls. The AUC for TNF- $\alpha$  was 0.86 (0.83–0.89). AUC, area under the curve; ROC, receiver operating characteristic; SLE, systemic lupus erythematosus.

were obtained for p-albumin, TNF- $\alpha$ , C3, IL-15, ESR, IL-10 and MIP-1 $\alpha$  ( $\rho=0.3$ ). If only patients with non-nephritis were considered (online supplementary table S2), the correlation between p-albumin and SLAM remained but weakened, while SLEDAI-2K lost significance. The correlation between SLEDAI-2K and IL-10 remained with the same strength. For PtGDA, the strongest correlation was obtained for p-albumin ( $\rho=-0.42$ ) followed by IL-6, TNF- $\alpha$ , MCP-1, IP-10 and hsCRP ( $\rho=0.3$ ). Including only patients without nephritis (online supplementary table S2), the correlations to PtGDA remained with similar strength for p-albumin ( $\rho=-0.36$ ), IL-6 ( $\rho=0.38$ ), MCP-1 ( $\rho=0.27$ ), IP-10 ( $\rho=0.26$ ) and hsCRP ( $\rho=0.29$ ), while it was decreased for TNF- $\alpha$  ( $\rho=0.14$ ). Correlations between the three investigated DA measurements were all moderate:

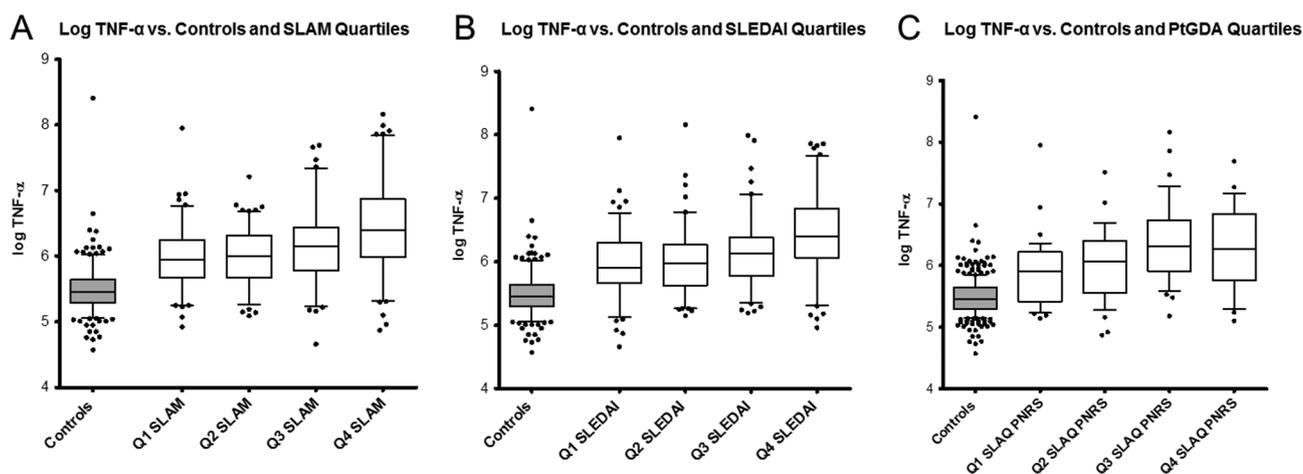
SLEDAI-2K vs SLAM with  $\rho=0.56$ , SLEDAI-2K vs PtGDA with  $\rho=0.44$  and SLAM vs PtGDA with  $\rho=0.58$ .

### Organ-specific measure of disease activity

We investigated if DA in specific organ systems was associated to our potential biomarkers. Comparing patients with active kidney involvement to patients with inactive or no renal involvement, p-albumin and TNF- $\alpha$  showed the highest association (table 3). Additionally, IL-16, anti-dsDNA and IL-10 were elevated in active nephritis (Bonferroni-corrected  $p<0.05$ ). Another measure of renal DA is the urinary (u-) albumin/creatinine ratio and it was as expected associated with active kidney involvement (table 3). This ratio is mainly relevant to measure in patients with nephritis, since patients with non-nephritis do not excrete albumin in the urine. The u-albumin/creatinine ratio was found to correlate to SLAM ( $\rho=0.22$ ) and SLEDAI-2K ( $\rho=0.30$ ) but not to PtGDA ( $\rho=0.12$ ,  $\rho=0.2$ ) (table 2) and in the non-nephritis subgroup, the correlation to SLEDAI-2K was not significant (online supplementary table S2). P-albumin and TNF- $\alpha$  levels correlated to u-albumin/creatinine ( $\rho=-0.39$  and  $\rho=0.28$ , respectively). In patients with joint involvement/active arthritis, IP-10, IL-1 $\alpha$ , IL-6, TNF- $\alpha$  and ESR were significantly increased (Bonferroni-corrected  $p<0.05$ ) compared with patients without active arthritis. IP-10 was associated to skin/mucosa involvement but the difference did not remain significant after Bonferroni correction.

### Patients with disease activity versus patients with inactive disease and controls

We defined inactive SLE as SLEDAI-2K=0 ( $n=115$ ), previously described as remission.<sup>15–17</sup> With this definition 26% had inactive disease. Of inactive patients, 41% ( $n=47$ ) did not take prednisolone at all and 78% ( $n=90$ ) were on a maximum prednisolone dose of 5 mg. Comparing inactive patients with the other patients (SLEDAI-2K>0,  $n=322$ ), we found that IL-15, TNF- $\alpha$ , IL-7, VEGF, IP-10, anti-dsDNA and ESR were higher and C4 and C3 lower



**Figure 3** The relationship of TNF- $\alpha$  to different measures of DA are shown: Log<sub>10</sub> TNF- $\alpha$  levels versus quartiles of (A) SLEDAI-2K, (B) SLAM and (C) PtGDA. DA, disease activity; PtGDA, patients' assessment of Global Disease Activity; SLAM, SLE Activity Measure; SLAQ, Systemic Lupus Activity Questionnaire; SLE, systemic lupus erythematosus; SLEDAI, SLE DA Index.

Table 3 Comparison between active and inactive disease in different organ systems

	Kidney*			Joints†			Skin/mucosa‡		
	Inactive (n=363) BILAG= C/D/E	Active (n=42) BILAG= A/B	P value§	Inactive (n=356) SLAM 24≤1	Active (n=74) SLAM 24≥2	P value§	Inactive (n=317) SLAM4+6=0	Active (n=109) SLAM4+6 ≥1	P value§
Cytokines¶									
INF-γ	11.5 (6.9–19.3)	18.5 (7.4–44.0)	0.03	11.5 (6.8–19.5)	12.3 (7.6–28.2)	0.2	10.9 (6.7–19.6)	13.0 (7.2–19.7)	0.09
IL-8	4.9 (3.1–8.4)	5.6 (3.2–13.6)	0.2	4.8 (3.0–8.4)	5.2 (3.4–10.1)	0.2	4.8 (3.0–8.2)	5.0 (3.3–9.8)	0.3
IL-15	2.9 (2.3–3.8)	4.2 (2.6–5.7)	<b>0.0007</b>	2.9 (2.3–4.1)	3.0 (2.4–4.0)	0.6	2.9 (2.3–4.1)	3.1 (2.4–3.9)	0.5
Eotaxin	132.6 (96.6–188.5)	144.1 (91.6–183.7)	0.6	133.5 (96.7–185.9)	129.2 (98.1–182.2)	0.9	131.8 (99.7–185.7)	134.8 (94.8–188.4)	0.7
MCP-1	105.2 (82.2–148.8)	137.5 (96.2–200.3)	0.007	106.7 (83.1–148.2)	119.2 (83.1–172.0)	0.2	108.8 (83.4–150.1)	111.0 (81.1–183.4)	0.6
MDC	880.9 (670.3– 1136.0)	677.0 (507.6–969.0)	0.002	843.9 (647.2– 1117.3)	814.4 (584.1– 1135.0)	0.5	838.5 (623.0–1111.4)	886.7 (662.8– 1173.5)	0.3
MIP-1β	68.9 (50.6–102.8)	94.6 (54.7–122.9)	0.05	69.7 (50.5–105.6)	79.3 (54.4–125.0)	0.1	73.8 (50.4–109.7)	67.0 (51.2–106.8)	0.6
IL-10	0.78 (0.50–1.38)	1.07 (0.83–3.38)	<b>0.002</b>	0.79 (0.49–1.40)	1.04 (0.64–1.93)	0.04	0.85 (0.51–1.42)	0.81 (0.52–1.78)	0.6
IL-6	1.09 (0.64–1.95)	1.96 (1.08–4.00)	<b>0.0004</b>	1.10 (0.66–2.01)	1.49 (0.82–3.30)	0.006	1.10 (0.67–2.07)	1.34 (0.68–2.29)	0.4
TNF-α	4.2 (3.0–5.8)	6.5 (5.5–10.9)	<b>6.4×10<sup>-9</sup></b>	4.4 (3.1–5.9)	5.6 (3.5–7.9)	0.006	4.4 (3.1–6.1)	4.6 (3.1–6.4)	0.4
IL-12/IL-23p40	172.0 (120.4–279.0)	214.6 (145.0–405.5)	0.01	178.9 (122.9–279.3)	196.7 (112.8–369.5)	0.2	180.4 (127.7–284.9)	185.9 (111.4–294.9)	0.7
IL-16	194.8 (145.4–259.6)	308.7 (228.2–413.6)	<b>1.7×10<sup>-7</sup></b>	208.6 (149.3–275.3)	235.7 (154.5–366.7)	0.04	216.6 (156.1–292.4)	186.0 (137.9–269.4)	0.03
IL-1α	6.1 (3.1–12.2)	8.4 (4.5–21.7)	0.04	5.6 (3.2–12.4)	9.0 (3.9–18.9)	0.005	6.4 (3.3–14.4)	5.3 (3.1–11.8)	0.5
IL-7	5.0 (3.1–8.5)	6.8 (4.4–9.2)	0.16	5.0 (3.0–8.8)	5.1 (3.2–8.1)	0.7	5.0 (3.0–8.7)	5.0 (3.2–8.4)	0.7
VEGF	74.8 (47.8–119.7)	86.1 (46.2–146.1)	0.27	73.2 (47.1–118.0)	91.4 (55.4–157.4)	0.046	74.1 (47.9–112.9)	81.6 (45.5–155.4)	0.5
Eotaxin-3	24.4 (19.3–34.2)	31.2 (19.0–35.4)	0.8	24.4 (19.4–34.9)	24.7 (19.1–34.6)	0.9	24.2 (19.4–35.6)	25.0 (18.9–34.6)	1.0
IP-10	739.6 (458.8– 1459.5)	1205.3 (393.5– 2428.9)	0.4	700.8 (426.5– 1311.3)	940.0 (605.3– 2566.7)	<b>0.0005</b>	705.2 (416.1–1370.6)	855.7 (529.2– 2123.0)	0.003
MCP-4	79.8 (54.4–129.3)	70.4 (48.3–95.3)	0.04	76.5 (53.2–122.4)	79.9 (53.8–126.2)	0.4	75.4 (53.6–119.3)	79.0 (51.9–129.0)	0.8
MIP-1α	20.3 (17.0–26.9)	29.5 (21.4–37.4)	<b>0.0009</b>	21.0 (17.1–27.2)	23.9 (17.8–32.0)	0.08	21.8 (17.3–30.8)	21.4 (17.2–24.9)	0.2
TARC	87.0 (53.3–151.0)	74.0 (41.0–132.1)	0.2	86.7 (51.3–153.0)	82.3 (55.9–131.6)	0.9	78.0 (51.2–138.7)	96.1 (54.4–160.1)	0.1
Standard clinical laboratory measurements¶									
ESR	18 (10–32)	30 (14–67)	<b>0.0001</b>	18 (10–32)	29 (13.2–38.8)	0.006	18 (10–33)	22 (13–39)	0.08
C4	0.15 (0.11–0.20)	0.13 (0.06–0.18)	0.05	0.15 (0.11–0.20)	0.13 (0.07–0.18)	0.02	0.15 (0.10–0.20)	0.15 (0.10–0.19)	0.6
C3	0.89 (0.74–1.05)	0.74 (0.55–0.87)	<b>0.0005</b>	0.88 (0.72–1.05)	0.82 (0.62–1.04)	0.1	0.88 (0.71–1.04)	0.87 (0.74–1.05)	0.8
P-albumin	39 (37–42)	31 (26–31)	<b>4.2×10<sup>-16</sup></b>	39 (36–42)	38 (34–41)	0.08	39 (36–42)	39 (35–41)	0.3
hsCRP	1.5 (0.6–4.7)	2.2 (1.1–8.9)	0.01	1.6 (0.6–4.9)	2.9 (0.8–7.4)	0.047	1.6 (0.6–4.9)	1.7 (0.8–6.2)	0.3

Continued

Table 3 Continued

	Kidney*			Joints†			Skin/mucosa‡		
	Inactive (n=363) BILAG=C/D/E	Active (n=42) BILAG=A/B	P value§	Inactive (n=356) SLAM 24≤1	Active (n=74) SLAM 24≥2	P value§	Inactive (n=317) SLAM4+6=0	Active (n=109) SLAM4+6≥1	P value§
Anti-dsDNA	4 (4–16)	22 (5–89)	<b>2.2×10<sup>-6</sup></b>	5 (4–17)	7 (4–49)	0.02	5 (4–18)	5 (4–26)	0.6
U-albumin/creatinine	0.8 (0.5–2.3)	75.4 (46.6–834.9)	<b>1.0×10<sup>-23</sup></b>	1.0 (0.5–5)	1.5 (0.7–12.2)	0.2	1.1 (0.5–6.7)	1.1 (0.5–4.9)	0.5

\*Renal BILAG A/B as measure of kidney involvement (active, n=42) vs renal BILAG=C/D/E as a measure of patients without active kidney involvement (n=363).

†Patients with SLAM item 24>1 (n=74) were defined as patients with joint involvement (active) and patients with SLAM item 24≤1 (n=356) were defined as patients with inactive joint involvement.

‡Skin/mucosal involvement if SLAM 4+SLAM 6>0 (active, n=109). No skin/mucosal involvement if SLAM 4+6 = 1 (n=317).

§P values (Mann-Whitney U-test) are reported and highlighted in bold if significant after Bonferroni correction for multiple testing.

¶Median (25% quantile–75% quantile). Concentrations of cytokines are reported in pg/mL. Standard clinical laboratory measurements reported as ESR in mm/hour, Complement factor C4 in g/L, Complement factor C3 in g/L, plasma albumin (p-albumin) in g/L, high-sensitivity (hs) CRP in mg/L, anti-dsDNA units and urinary (u) albumin/creatinine ratio in mg/mmol.

BILAG, British Isles Lupus Assessment Group, Renal index; ESR, erythrocyte sedimentation rate; SLAM, SLE Activity Measure; SLE, systemic lupus erythematosus.

in patients with DA ( $p<0.001$ ). P-albumin did not reach this level of significance after Bonferroni correction comparing active and inactive disease ( $P=0.008$ , not corrected for multiple testing). Interestingly, IL-7 and VEGF did not differ when comparing inactive patients with controls. All other investigated variables except MDC, IL-16, IL-1 $\alpha$  and Eotaxin-3 differed between inactive patients and controls (online supplementary table S3). Five of the inactive patients (SLEDAI-2K=0) also had SLAM=0. Only eight patients in the cohort had SLAM=0.

## DISCUSSION

The major finding in this study is that circulating levels of TNF- $\alpha$  outperform a set of 26 investigated traditional and potential SLE biomarkers as a discriminator between patients with SLE and controls. Furthermore, TNF- $\alpha$  levels correlate with DA (SLAM, SLEDAI and PtGDA) and is in particular associated with active nephritis. Together with previous reports our results demonstrate that TNF- $\alpha$  plays an important role in SLE and deserves attention as a clinically useful biomarker.

Though initial reports were inconsistent, there are now a large number of studies, which reliably demonstrate that high levels of circulating TNF- $\alpha$  are associated with SLE,<sup>10 12 18–21</sup> regardless of ethnicity.<sup>22</sup> Many positive correlations between TNF- $\alpha$  and higher DA have been reported<sup>10 12 23 24</sup> and these together with our results may still be underestimations since most active patients are taking prednisolone and other immunosuppressive treatments, which are likely to have a negative impact on cytokine levels.<sup>25</sup> We previously noted that TNF- $\alpha$  alone, and combined with its highly correlated soluble receptors (TNFR1 and TNFR2),<sup>26</sup> is a good measure/biomarker of DA in SLE.<sup>10 12</sup> Others have reported similar results.<sup>27</sup> Cigni *et al* found the highest levels of TNF- $\alpha$  in patients with newly diagnosed active and untreated SLE,<sup>19</sup> while Munroe *et al* demonstrated that high levels of TNF- $\alpha$  and

its two soluble receptors precede disease flares by 6–12 weeks.<sup>28</sup> In a smaller SLE cohort (n=45), investigating five cytokines by the MSD assay, correlations between TNF- $\alpha$  and SLEDAI were similar to our observations.<sup>12</sup> The novelty of this study is that we can demonstrate a supreme performance of TNF- $\alpha$  in a competitive context including a large group of patients and matched population controls, many new and traditional biomarkers and three DA indices, one of which is patient reported. TNF- $\alpha$  blockade in SLE has been reported in smaller open label studies and case reports and is still a controversial issue. Concerns regarding induction of autoantibodies and infections remain, nevertheless many patients with SLE have improved following such therapy.<sup>29 30</sup>

Importantly, our results also demonstrate that p-albumin performed almost as well as TNF- $\alpha$ , demonstrating inverse correlation to both SLAM and SLEDAI and even stronger to PtGDA. Yip *et al* previously reported negative correlations between albumin levels and DA in SLE, and in accordance with our results correlations were strongest among patients with lupus nephritis, but also present in patients with non-nephritis SLE.<sup>31</sup> These very useful findings are, however, not yet widely recognised or highlighted and they are not used in any of the commonly used DA indices.<sup>3–6</sup> The absolute differences in albumin levels may, however, be small, especially in patients with non-nephritis as demonstrated in online supplementary table S2. To further evaluate albumin as a measure of DA, we suggest that levels should be followed in longitudinal studies, that is, using each individual as its own control. Albumin, a negative acute-phase protein and a marker of inflammation,<sup>32</sup> correlates with degree of illness<sup>33</sup> in many settings, thus not specifically with SLE-related DA. In renal disease, albumin levels are indirect reflections of renal loss through proteinuria. Albumin is also routinely used to evaluate protein and energy/nutritional status.<sup>34</sup> Though unspecific, it is important to highlight that

p-albumin, an inexpensive routine laboratory analysis performed by simple instrumentation with a fast readout, is a good biomarker for SLE-related DA, especially in patients with nephritis. P-albumin also reflected PtGDA well both in patients with nephritis and non-nephritis. In contrast, several current DA indices require specialised laboratories where analyses are costly and slow, for example, anti-dsDNA antibodies, C3 and C4. In a global perspective where resources are often limited, our observation is highly relevant in order to obtain equal health and treatment for patients with SLE.

Interestingly, IL-15, a regulator of T-cells and natural killer cells, correlated with SLAM, SLEDAI and was associated with renal DA. Furthermore, IL-15 was a good discriminator between controls, inactive and active SLE. Aringer *et al* previously reported high levels of IL-15<sup>35</sup> and recently urinary levels of IL-15 were reported to discriminate active lupus nephritis from active SLE with other manifestations.<sup>36</sup> To our knowledge, circulating IL-15 has not previously been investigated in a large SLE cohort. IL-16 is mainly a CD8+ T cell cytokine, which induces chemotaxis of CD4+ T cells and monocytes. We confirm previous reports that IL-16 levels are higher in SLE than controls.<sup>37,38</sup> While correlations with overall DA were modest, we noted strong associations with active renal disease.

We evaluated DA in three different organ systems: kidneys, joints and skin/mucosa. We observed increased levels of TNF- $\alpha$ , IL-16, anti-dsDNA and IL-10 and lower levels of p-albumin in patients with active kidney disease as compared with other patients with lupus. The association between renal BILAG A/B and high TNF- $\alpha$  levels is supported by previously documented local upregulation of genes, single-cell RNA and production of TNF- $\alpha$  in active renal disease. TNF- $\alpha$  was in these studies located in tubular cells and along glomeruli.<sup>39–41</sup> Furthermore, high circulating levels of TNF- $\alpha$  together with IL-17/IL-23 and INF- $\gamma$  indicated poor response to traditional treatment in lupus nephritis.<sup>42,43</sup> Preserved P-albumin was recently reported to predict favourable outcome after 4 years in patients with biopsy proven nephritis.<sup>44</sup> Interestingly, IP-10 was upregulated in active arthritis and in skin/mucosa manifestations, but not in active nephritis. We confirm our previous observation that IP-10 is associated with high DA, arthritis and musculoskeletal symptoms<sup>45</sup> and those of Kong *et al* who reported associations between IP-10 and both general and mucocutaneous DA.<sup>46</sup> Joint and skin/mucosa manifestations are likely to affect patients' perception of disease and IP-10 also correlated more strongly to PtGDA than to SLAM and SLEDAI. IFN- $\alpha$  was not included in the Human Cytokine 30-plex assay and was therefore not analysed. However, we recently investigated a subset of this cohort with a pan-IFN- $\alpha$  ELISA assay, demonstrating that high levels of IFN- $\alpha$  were associated with active mucocutaneous disease.<sup>45</sup> Furthermore, we observed higher TNF- $\alpha$ , IL-6, IL-1 $\alpha$  and ESR in patients with joint activity. Taken together the pattern of biomarkers seems to differ between organ involvements.

A great advantage with this study is that different instruments have been used to measure DA. Agreements between these measures of DA have been previously studied<sup>47</sup> and the correlations are commonly about  $\rho=0.6$ , thus in line with our findings. All frequently used DA indices require formal training and substantial time for a proper assessment.<sup>1</sup> Nevertheless, subjective perceptions definitely influence the results. Previous studies have pointed out the importance to also incorporate elements of patients' perception when assessing DA.<sup>1</sup> An important advantage with patients' assessments of DA is that it is easy<sup>14</sup> and inexpensive to collect in clinical practice. In concordance with this discussion, it is a great advantage that we can compare our results with PtGDA from a subset of our patients. Interestingly, PtGDA correlated best with p-albumin, IL-6, TNF- $\alpha$ , MCP-1 and IP-10, all of which also correlated to SLAM and SLEDAI. Thus, these cytokines seem to be pivotal in organ manifestations with impact on patients' health perception. On the other hand, IL-15, MIP-1 $\alpha$ , ESR correlated poorly with PtGDA, despite correlations with SLAM and SLEDAI-2K, suggesting that these cytokines reflect disease processes which are difficult for patients to assess. Renal activity is one such manifestation, and PtGDA had low correlations with IL-16 and MIP-1 $\alpha$ , while these cytokines were associated with active nephritis.

The large size and detailed clinical information in this study are strengths, while the cross-sectional design is a weakness and longitudinal confirmation is needed to evaluate the predictive power of investigated biomarkers. However, others have reported similar associations with TNF- $\alpha$  in longitudinal studies supporting our results of TNF- $\alpha$  as a marker of DA<sup>28</sup> and p-albumin.<sup>31,44</sup> Another weakness with this study is that mainly European Caucasian patients/controls were included and results can thus not be generalised to other ethnicities. Furthermore, the specificity of the suggested biomarkers has not been studied in relation to other diseases.

Based on a large and well-characterised cohort, a sizeable set of biomarkers and DA measured, we demonstrate that TNF- $\alpha$  and p-albumin are supreme discriminators between patients with SLE and controls and they also correlate with  $\rho \geq 0.3$  to all three DA indices. The reproducibility of TNF- $\alpha$  data in several previous cohorts and the routine use of p-albumin in the clinic highlight the robustness of the suggested biomarkers. We suggest that TNF- $\alpha$  and p-albumin can be used in the clinic at a reasonable cost as useful biomarkers for diagnostic and surveillance purposes in SLE.

**Acknowledgements** We would like to acknowledge patients and participants in the study. We are grateful to Eva Jemseby for management of blood samples and to Jill Gustafsson and Sonia Möller for coordination and blood sampling. We thank Ola Börjesson for help with inclusion of patients.

**Contributors** HI: study design, data analysis and manuscript writing. SE: cytokine measurements, manuscript writing. SP: PtGDA data collection and analysis, manuscript writing. JTG: clinical data, manuscript writing/approval. AZ: clinical data, manuscript writing/approval. MK: clinical data, manuscript writing/approval. VO: clinical data, manuscript writing/approval. P-JJ: study design, manuscript writing/

approval. IG: SLE cohort responsible, manuscript writing. ES: study design, SLE cohort responsible, data analysis, manuscript writing.

**Funding** This study was supported by the AstraZeneca-Karolinska Institutet Joint Research Program in Translational Science, Swedish Research Council, Stockholm County Council (ALF), Swedish Heart-Lung foundation, King Gustaf Vs 80th Birthday Fund, Swedish Rheumatism Association, Swedish Society of Medicine, The Åke Wiberg Foundation, Karolinska Institutet's Foundations and The Foundation in memory of Clas Groschinsky.

**Competing interests** SE is an employee of AstraZeneca.

**Patient consent** Obtained.

**Ethics approval** The Local Ethics Committee of the Karolinska University Hospital/Karolinska Institutet in Stockholm approved the study.

**Provenance and peer review** Not commissioned; externally peer reviewed.

**Data sharing statement** There are no additional unpublished data available.

**Open Access** This is an Open Access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

© Article author(s) (or their employer(s) unless otherwise stated in the text of the article) 2018. All rights reserved. No commercial use is permitted unless otherwise expressly granted.

## REFERENCES

- Mikdashi J, Nived O. Measuring disease activity in adults with systemic lupus erythematosus: the challenges of administrative burden and responsiveness to patient concerns in clinical research. *Arthritis Res Ther* 2015;17:183.
- Bombardier C, Gladman DD, Urowitz MB, et al. Derivation of the SLEDAI. A disease activity index for lupus patients. *Arthritis & Rheumatism* 1992;35:630–40.
- Gladman DD, Ibañez D, Urowitz MB. Systemic lupus erythematosus disease activity index 2000. *J Rheumatol* 2002;29:288.
- Liang MH, Socher SA, Larson MG, et al. Reliability and validity of six systems for the clinical assessment of disease activity in systemic lupus erythematosus. *Arthritis Rheum* 1989;32:1107–18.
- Mosca M, Bencivelli W, Vitali C, et al. The validity of the ECLAM index for the retrospective evaluation of disease activity in systemic lupus erythematosus. *Lupus* 2000;9:445–50.
- Hay EM, Bacon PA, Gordon C, et al. The BILAG index: a reliable and valid instrument for measuring clinical disease activity in systemic lupus erythematosus. *Q J Med* 1993;86:447.
- Karlson EW, Daltroy LH, Rivest C, et al. Validation of a Systemic Lupus Activity Questionnaire (SLAQ) for population studies. *Lupus* 2003;12:280–6.
- Ohl K, Tenbrock K. Inflammatory cytokines in systemic lupus erythematosus. *J Biomed Biotechnol* 2011;2011:1–14.
- Adhya Z, Borozdenkova S, Karim MY. The role of cytokines as biomarkers in systemic lupus erythematosus and lupus nephritis. *Nephrol Dial Transplant* 2011;26:3273–80.
- Svenungsson E, Gunnarsson I, Fei GZ, et al. Elevated triglycerides and low levels of high-density lipoprotein as markers of disease activity in association with up-regulation of the tumor necrosis factor alpha/tumor necrosis factor receptor system in systemic lupus erythematosus. *Arthritis Rheum* 2003;48:2533–40.
- Gröndal G, Gunnarsson I, Rönnelid J, et al. Cytokine production, serum levels and disease activity in systemic lupus erythematosus. *Clin Exp Rheumatol* 2000;18:565–70.
- McCarthy EM, Smith S, Lee RZ, et al. The association of cytokines with disease activity and damage scores in systemic lupus erythematosus patients. *Rheumatology* 2014;53:1586–94.
- Tan EM, Cohen AS, Fries JF, et al. The 1982 revised criteria for the classification of systemic lupus erythematosus. *Arthritis Rheum* 1982;25:1271–7.
- Pettersson S, Svenungsson E, Gustafsson J, et al. A comparison of patients' and physicians' assessments of disease activity using the Swedish version of the Systemic Lupus Activity Questionnaire. *Scandinavian Journal of Rheumatology* 2017:1–10.
- Gkw L, Petri M. Assessment of systemic lupus erythematosus. *Clinical and experimental rheumatology* 2005;235(Suppl 39):S120.
- Polachek A, Gladman DD, Su J, et al. Defining low disease activity in systemic lupus erythematosus: Low disease activity in SLE. *Arthritis Care & Research* 2016.
- Zen M, Iaccarino L, Gatto M, et al. Prolonged remission in Caucasian patients with SLE: prevalence and outcomes. *Ann Rheum Dis* 2015;74:2117–22.
- Aringer M, Smolen JS. The role of tumor necrosis factor-alpha in systemic lupus erythematosus. *Arthritis Res Ther* 2008;10:202.
- Cigni A, Pileri PV, Faedda R, et al. Interleukin 1, interleukin 6, interleukin 10, and tumor necrosis factor  $\alpha$  in active and quiescent systemic lupus erythematosus. *J Investig Med* 2014;62:825–9.
- Studnicka-Benke A, Steiner G, Petera P, et al. Tumor necrosis factor alpha and its soluble receptors parallel clinical disease and autoimmune activity in systemic lupus erythematosus. *Br J Rheumatol* 1996;35:1067–74.
- Svenungsson E, Fei GZ, Jensen-Urstad K, et al. TNF-alpha: a link between hypertriglyceridaemia and inflammation in SLE patients with cardiovascular disease. *Lupus* 2003;12:454–61.
- Weckerle CE, Mangale D, Franek BS, et al. Large-scale analysis of tumor necrosis factor  $\alpha$  levels in systemic lupus erythematosus. *Arthritis Rheum* 2012;64:2947–52.
- Davas EM, Tsirogianni A, Kappou I, et al. Serum IL-6, TNFalpha, p55 srTNFalpha, p75srTNFalpha, srIL-2alpha levels and disease activity in systemic lupus erythematosus. *Clin Rheumatol* 1999;18:17–22.
- Postal M, Lapa AT, Sinicato NA, et al. Depressive symptoms are associated with tumor necrosis factor alpha in systemic lupus erythematosus. *J Neuroinflammation* 2016;13:5.
- Song L, Qiu F, Fan Y, et al. Glucocorticoid regulates interleukin-37 in systemic lupus erythematosus. *J Clin Immunol* 2013;33:111–7.
- Heilig B, Fiehn C, Brockhaus M, et al. Evaluation of soluble tumor necrosis factor (TNF) receptors and TNF receptor antibodies in patients with systemic lupus erythematosus, progressive systemic sclerosis, and mixed connective tissue disease. *J Clin Immunol* 1993;13:321–8.
- Aderka D, Wysesbeek A, Engelmann H, et al. Correlation between serum levels of soluble tumor necrosis factor receptor and disease activity in systemic lupus erythematosus. *Arthritis Rheum* 1993;36:1111–20.
- Munroe ME, Vista ES, Guthridge JM, et al. Proinflammatory adaptive cytokine and shed tumor necrosis factor receptor levels are elevated preceding systemic lupus erythematosus disease flare. *Arthritis Rheumatol* 2014;66:1888–99.
- Aringer M, Smolen JS. Therapeutic blockade of TNF in patients with SLE-promising or crazy? *Autoimmun Rev* 2012;11:321–5.
- Cortés-Hernández J, Egri N, Vilardell-Tarrés M, et al. Etanercept in refractory lupus arthritis: An observational study. *Semin Arthritis Rheum* 2015;44:672–9.
- Yip J, Aghdassi E, Su J, et al. Serum albumin as a marker for disease activity in patients with systemic lupus erythematosus. *J Rheumatol* 2010;37:1667–72.
- Dominguez de Villota E, Mosquera JM, Rubio JJ, et al. Association of a low serum albumin with infection and increased mortality in critically ill patients. *Intensive Care Med* 1980;7:19–22.
- Van Hemelrijck M, Harari D, Garmo H, et al. Biomarker-based score to predict mortality in persons aged 50 years and older: a new approach in the Swedish AMORIS study. *Int J Mol Epidemiol Genet* 2012;3:66.
- Friedman AN, Fadem SZ. Reassessment of albumin as a nutritional marker in kidney disease. *J Am Soc Nephrol* 2010;21:223–30.
- Aringer M, Stummvoll GH, Steiner G, et al. Serum interleukin-15 is elevated in systemic lupus erythematosus. *Rheumatology* 2001;40:876–81.
- Landolt-Marticorena C, Prokopec SD, Morrison S, et al. A discrete cluster of urinary biomarkers discriminates between active systemic lupus erythematosus patients with and without glomerulonephritis. *Arthritis Res Ther* 2016;18:218.
- Lee S, Kaneko H, Sekigawa I, et al. Circulating interleukin-16 in systemic lupus erythematosus. *Br J Rheumatol* 1998;37:1334–7.
- Lard LR, Roep BO, Verburgh CA, et al. Elevated IL-16 levels in patients with systemic lupus erythematosus are associated with disease severity but not with genetic susceptibility to lupus. *Lupus* 2002;11:181–5.
- Malide D, Russo P, Bendayan M. Presence of tumor necrosis factor alpha and interleukin-6 in renal mesangial cells of lupus nephritis patients. *Hum Pathol* 1995;26:558–64.
- Herrera-Esparza R, Barbosa-Cisneros O, Villalobos-Hurtado R, et al. Renal expression of IL-6 and TNFalpha genes in lupus nephritis. *Lupus* 1998;7:154–8.
- Der E, Suryawanshi H, Ranabothu S, et al. Kidney and Skin Single-Cell RNA Sequencing in Lupus Nephritis Provides Mechanistic

- Insights and Novel Potential Biomarkers. *Arthritis Rheumatology* 2017;69(Suppl 10).
42. Zickert A, Amoudruz P, Sundström Y, *et al.* IL-17 and IL-23 in lupus nephritis - association to histopathology and response to treatment. *BMC Immunol* 2015;16:7.
  43. Parodis I, Ding H, Zickert A, *et al.* Serum soluble tumour necrosis factor receptor-2 (sTNFR2) as a biomarker of kidney tissue damage and long-term renal outcome in lupus nephritis. *Scand J Rheumatol* 2017;46:263–72.
  44. Domingues V, Bornkamp N, Levinson B, *et al.* Serum Albumin at 12 Months Post Biopsy Has Excellent Sensitivity and Specificity for Favorable 4 Year Renal Outcome in Lupus Nephritis (LN) [abstract]. *Arthritis Rheumatology* 2017;69(Suppl 10).
  45. Oke V, Brauner S, Larsson A, *et al.* IFN-  $\gamma$  with Th17 axis cytokines and IFN- $\alpha$  define different subsets in systemic lupus erythematosus (SLE). *Arthritis Res Ther* 2017;19:139.
  46. Kong KO, Tan AW, Thong BY, *et al.* Enhanced expression of interferon-inducible protein-10 correlates with disease activity and clinical manifestations in systemic lupus erythematosus. *Clin Exp Immunol* 2009;156:134–40.
  47. Uribe AG, Vilá LM, McGwin G, *et al.* The Systemic Lupus Activity Measure-revised, the Mexican Systemic Lupus Erythematosus Disease Activity Index (SLEDAI), and a modified SLEDAI-2K are adequate instruments to measure disease activity in systemic lupus erythematosus. *J Rheumatol* 2004;31:1934.
  48. Gladman D, Ginzler E, Goldsmith C, *et al.* The development and initial validation of the Systemic Lupus International Collaborating Clinics/American College of Rheumatology damage index for systemic lupus erythematosus. *Arthritis Rheum* 1996;39:363–9.
  49. Franklyn K, Lau CS, Navarra SV, *et al.* Definition and initial validation of a Lupus Low Disease Activity State (LLDAS). *Ann Rheum Dis* 2016;75:1615–21.