Clinical phenotype associations with various types of anti-dsDNA antibodies in patients with recent onset of rheumatic symptoms. Results from a multicentre observational study

Michele Compagno, Ole P Rekvig, Anders A Bengtsson, Gunnar Sturfelt, Niels H Heegaard, Andreas Jønsen, Rasmus Sleimann Jacobsen, Gro Ø Eilertsen, Christopher G Fenton, Lennart Truedsson, Johannes C Nossent, Søren Jacobsen

ABSTRACT

Despite anti-dsDNA antibodies constitute a wide range of specificities, they are considered as the hallmark for systemic lupus erythematosus (SLE).

Objective: To identify clinical phenotypes associated with anti-dsDNA antibodies, independently of any clinical diagnoses.

Methods: Patients with recent onset of any rheumatic symptoms were screened for antinuclear antibodies (ANA). All ANA-positive and matching ANA-negative patients were examined, and their clinical phenotypes were registered, using a systematic chart formulated after consensus between the participating centres. All patients were tested for different anti-dsDNA antibody specificities with assays habitually used in each participating laboratory. Crithidia Luciliae Immuno Fluorescence Test (CLIFT) was performed three times (with two different commercial kits); solid and solution phase ELISA were performed four times. Associations between clinical phenotypes and results of anti-dsDNA assays were evaluated by linear regression analysis (LRA) and principal component analysis (PCA).

Results: Totally, 292 ANA-positive and 292 matching ANA-negative patients were included in the study. A full dataset for statistical analysis was obtained in 547 patients. Anti-dsDNA antibodies were most frequently detected by ELISA. LRA showed that overall positivity of anti-dsDNA antibodies was associated with proteinuria and pleuritis. Alopecia was significantly associated with CLIFT-positivity. Besides confirming the same findings, PCA showed that combined positivity of CLIFT and ELISA was also associated with lymphopenia.

Conclusions: Our results show that different anti-dsDNA antibody specificities are associated with nephropathy, pleuritis, alopecia and lymphopenia, regardless of the diagnosis. It may challenge the importance of anti-dsDNA antibodies as a diagnostic hallmark for SLE.

KEY MESSAGES

- In patients with recent onset of rheumatic symptoms, the assessment of anti-dsDNA antibodies with different techniques results in a considerable discrepancy of outcomes and of correlations to various clinical and biochemical manifestations.
- Anti-dsDNA antibodies are associated with presence of proteinuria, regardless of clinical diagnosis, outcome of ANA screening and laboratory method used for the assessment of anti-dsDNA antibodies.
- In distinct subgroups of patients, anti-dsDNA antibodies are also variously associated with presence of other clinical manifestations, such as haematuria, leukopenia, pleuritis and alopecia.

Systemic lupus erythematosus (SLE) is a systemic autoimmune disease with unknown etiology. Whether SLE represents one disease entity or is a continuous overlap of aetiologically unrelated organ manifestations is not established.

This is particularly challenging when attempting to determine biomarkers for SLE. Anti-double-stranded DNA (dsDNA) antibodies are regarded as fairly specific for SLE.1-3 B-cell-mediated and T-cell-mediated autoimmunity to the individual components of nucleosomes are considered important in establishing a diagnosis,4,7 but the pathogenic and diagnostic roles played by anti-dsDNA and other antibodies are still debated.7,8

Antibodies to dsDNA may have a direct pathogenic effect in lupus nephritis,9 lupus dermatitis10,11 and possibly also in certain...
aspects of cerebral lupus.12 How anti-dsDNA antibodies relate to the rest of the clinical components of current classification criteria58 remains to be determined.

When emphasising anti-dsDNA antibodies as a central biomarker in SLE, it is important to perceive that these antibodies basically are not representing a homogenous antibody population.15–16 Growing insight into the different possible mechanisms of production of antibodies specifically binding to dsDNA17–24 challenges the notion of a specific relationship between all anti-dsDNA antibodies per se with SLE. Which and how anti-dsDNA antibodies are pathogenic has also been questioned.9 25–38

We aim to explore in the present investigation whether the positivity of anti-dsDNA antibodies is a biomarker indicating presence of defined clinical phenotypes, such as, for example, arthropathy or nephropathy or serositis, rather than a defined diagnosis, such as SLE.

The current literature is mainly composed of investigations of patients with established SLE and other defined and classified diagnoses. Studies concerning the association of anti-dsDNA antibodies with clinical manifestations in unselected patients with early onset of rheumatic symptoms are scarce.

The main purpose of this study is to correlate the presence in serum of anti-dsDNA antibodies with individual clinical manifestations and laboratory variables. We intend to use an unbiased approach that mirrors an ordinary clinical setting, where the physician is challenged to make the right diagnosis in patients with newly developed rheumatic manifestations, based upon clinical signs and symptoms, with the support of various diagnostic procedures, including laboratory tests. This approach also allows us to perform a comparison between serum levels of anti-dsDNA antibodies obtained in different laboratories with different methods.

**METHODS AND PATIENTS**

**Patients**

Consecutive patients with recent onset of suspected rheumatic disorder, referred for the first time to the participating rheumatologic units (Rigshospitalet in Copenhagen, Denmark; University Hospital in Tromsø, Norway; University Hospital in Lund, Sweden) were recruited to the study between February 2003 and December 2007.

Exclusion criteria were: established autoimmune disease, treatment with any biological drug, corticosteroids (equivalent Prednisolon >20 mg/day), immune-modulating, immunosuppressive or cytostatic drugs. Patients who previously had been examined by a rheumatologist, and patients unable to fully collaborate in the study (not confident with the language, actual cognitive, speech, hearing or memory impairment) were also excluded.

The patients were examined by a rheumatologist who made an initial working clinical diagnosis, based on anamnesis, symptoms, physical examination and laboratory test results.

All the patients were screened for antinuclear antibodies (ANA) by local testing. All the ANA-positive patients and the same amount (1:1 ratio) of randomly selected sex-matched and age-matched ANA-negative patients built together a nested cohort, which underwent further clinical and laboratory assessments. A systematic chart including clinical data and routine laboratory variables (see online supplementary table S1) was completed, ensuring a common trunk of data on which the clinical phenotypes of the patients could be characterised. Aliquots of serum samples from all patients were collected and sent to each centre for simultaneous parallel analysis of anti-dsDNA antibodies by several different assays, as described below. All clinical data were collected without knowledge of the results of the anti-dsDNA testing.

**Ethics**

All patients entered the study after giving informed written consent. The participating centres performed the study according to the approval from the local ethics committees.

**Clinical phenotype description**

Consensus as to the content of the clinical and laboratory dataset in the systematic chart (see online supplementary table S1) was obtained through a Delphi-like process, spanning over four meetings. A uniform definition of the final clinical dataset was assured by agreeing on the definitions of the various manifestations prior to the study (see online supplementary table S2). Manifestations were recorded as being absent ever, ongoing/active, previous/inactive or unknown and the date any manifestations first appeared was noted. In the present study, calculations were based on the presence ever of a manifestation.

**Detection of ANA**

In the various centres, respective current routine methodology was used for detection of ANA. In Copenhagen and Lund, screening for ANA was performed by indirect immunofluorescence (IIF) technique. In Copenhagen, HEP-2 cells (ImmunoConcepts, Sacramento, California, USA) and patient sera in dilution 1/160 were used together with an FITC-labelled antihuman IgG conjugate (DAKO Denmark A/S, Glostrup, Denmark). The diagnostic ANA titre was 1:160 as established by determination of the 95th percentile for negativity in healthy blood donors. In Lund, HEP-2 or HEP-20-10 cells (Euroimmun, Lübeck, Germany) and patient sera in dilution 1/400 were used which corresponded to detection of ANA with homogeneous pattern at 14 IU/mL (WHO reference serum 66/233). The diagnostic ANA titre was 1:400 as established by determination of the 96.5th percentile for negativity in healthy blood donors. In Tromsø, the detection of ANA was performed with the ELISA Varelisa ReCombi ANA Screen (Pharmacia
Diagnostics, Freiburg, Germany). This is an indirect non-competitive enzyme immunoassay for the qualitative and semiquantitative determination of eight preselected ANA (dsDNA, Sm (B,B’,D), ribo-nucleoprotein (68 kDa, A, C), SS-A/Ro(52 and 60 kDa), SS-B/La, Scl-70, centromere and Jo-1) in serum or plasma. The assay was performed as recommended by the manufacturer, while validation and determination of cut-off values were performed in accordance with an internal and external quality assessment programme, with validation against both controls and other disease categories.

Detection of anti-dsDNA antibodies
Without knowledge of any clinical information, all the patients included in the nested cohort had IgG anti-dsDNA antibodies determined by different indirect immunofluorescence (IIF) tests and immunoassays available at the participating centres (table 1).

Immunofluorescent tests
Presence of anti-dsDNA antibodies was assessed in all the three participating laboratories by Crithidia Luciliae Immunofluorescence Test (CLIFT) with two different commercial kits, according to the manufacturers’ instructions. By using a fluorescence microscope, the results were based on the fluorescence intensity and categorised as negative or positive.

Immunooassays
The determination of anti-dsDNA antibodies by the EliA test and the Varelisa were performed according to the manufacturer’s instructions.

The solution phase anti-dsDNA ELISA (SPADE) is previously described in detail. It measures antibody binding to dsDNA in solution using biotinylated dsDNA. Biotinylation of the pUC18 DNA (1 μg/μL H₂O) was carried out as recommended by the manufacturer (Pierce Chemical Company, Rockford, Illinois, USA). SPADE was performed by mixing 0.5 μg of pUC18 DNA (1 μg/μL H₂O) with serially diluted serum samples (solution phase step). After incubation for 30 min, the mixtures were added to microtitre plates (Nunc MaxiSorp, Nunc, Denmark) coated overnight at 37°C with 50 μL streptavidin (5 μg/mL PBS). After incubation for 30 min, the plates were washed and incubated with horseradish peroxidase conjugated anti-human Fcγ antibodies (Sigma–Aldrich, St Louis, Missouri, USA) in phosphate buffered saline with tween 20. The reaction was developed by adding o-phenylenediamine dihydrochloride, and stopped by 1 M HCl. The reaction was read at 490 nm.

Statistical analysis
After finalising data retrieval, all data were registered in a common database accessible for all centres. Statistical analyses using SPSS statistics V20.0 software (IBM) included contingency table analysis and binary logistic regression analysis. OR and corresponding 95% CIs were initially calculated as crude values using dichotomised anti-dsDNA results as the dependent variable and each dichotomised clinical manifestation registered in our database as explanatory variables. OR and corresponding 95% CI were then adjusted, using all the crude significant associations as covariates.

Principal component analysis (PCA) using R (http://www.R-project.org) included clinical phenotype data as well as positivity in the ANA test, any of the CLIFTs and any of the ELISAs. The PCA results were presented as biplots with variable vectors (arrows) indicating by direction which variables had the highest degree of covariation and influence, positive or negative, in discriminating the patients.

RESULTS
Altogether, 1073 patients were recruited from February 2003 to December 2007. Among these patients, 292 were found to be ANA positive by local testing. From the remaining ANA-negative patients, each centre in a 1:1 ratio randomly selected sex-matched and age-matched patients, reaching a total of 292 ANA-negative control patients. The resulting nested cohort of 584 patients (138 from

<table>
<thead>
<tr>
<th>Name</th>
<th>Methodology</th>
<th>Antigen</th>
<th>Reference interval</th>
<th>Manufacturer</th>
<th>Centre</th>
</tr>
</thead>
<tbody>
<tr>
<td>CLIFT 1</td>
<td>IIF</td>
<td>Crithidia luciliae kinetoplast</td>
<td>&lt;titre 10</td>
<td>ImmunoConcept</td>
<td>CPH</td>
</tr>
<tr>
<td>CLIFT 2</td>
<td>IIF</td>
<td>Crithidia luciliae kinetoplast</td>
<td>&lt;titre 10</td>
<td>ImmunoConcept</td>
<td>Tromsø</td>
</tr>
<tr>
<td>CLIFT 3</td>
<td>IIF</td>
<td>Crithidia luciliae kinetoplast</td>
<td>&lt;titre 10</td>
<td>Euroimmune</td>
<td>Lund</td>
</tr>
<tr>
<td>EliA</td>
<td>Solid phase ELISA</td>
<td>Recombinant plasmid dsDNA</td>
<td>&lt;10 IU/mL</td>
<td>Phadia</td>
<td>Tromsø</td>
</tr>
<tr>
<td>SPADE</td>
<td>Solution phase ELISA</td>
<td>Biotinylated plasmid dsDNA and biotinylated, S1 nucleaseme human dsDNA</td>
<td>&lt;1 AU/mL</td>
<td>Phadia</td>
<td>Tromsø</td>
</tr>
<tr>
<td>Varelisa 1</td>
<td>Solid phase ELISA</td>
<td>Recombinant plasmid dsDNA</td>
<td>≤55 IU/mL</td>
<td>Phadia</td>
<td>Tromsø</td>
</tr>
<tr>
<td>Varelisa 2</td>
<td>Solid phase ELISA</td>
<td>Recombinant plasmid dsDNA</td>
<td>≤35 IU/mL</td>
<td>Phadia</td>
<td>CPH</td>
</tr>
</tbody>
</table>

AU, arbitrary units; CLIFT, Crithidia Luciliae Immunofluorescence Test; CPH, Copenhagen; IIF: indirect immunofluorescence technique; IU, international units; SPADE, solution phase anti-dsDNA ELISA.
Clinical and biochemical manifestations in ANA positive and ANA-negative patients

A full serological dataset was obtained for 547 patients, 288 ANA positive (78.5% females, median age 51.8 years, range 15.4–83.7 years) and 259 ANA negative (83.4% females, median age 51.1 years, range 15.9–84.2 years). The remaining 37 patients, 4 ANA positive (75% females, median age 64 years, range 49–77 years) and 33 ANA negative (76% females, median age 59 years, range 22–81 years) withdrew from the study because of technical and logistic problems that hampered the collection of a full serological dataset.

The initial clinical diagnoses of the patients based on the conclusion of the examining rheumatologist are listed in table 2.

The most prevalent American College of Rheumatology (ACR) classification criteria for SLE,⁵ ⁴⁰ and the most relevant remaining clinical and biochemical manifestations are listed in table 3. Arthritis, photo-sensitivity, oral/nasal ulcers, haematuria, and proteinuria were the five most prevalent SLE specific manifestations. Malar rash was the only clinical variable significantly associated with the presence of ANA.

Arthralgia, morning joint stiffness, Raynaud’s phenomenon, headache, and subjective xerostomia were the five most prevalent manifestations among those not included in ACR classification criteria. Raynaud’s phenomenon, headache, puffy fingers and affective disorders were significantly more prevalent in the presence of ANA, whereas psoriasis was significantly more prevalent in ANA-negative patients.

### Table 2 Clinical diagnoses formulated by the examining rheumatologists in the participating centre

<table>
<thead>
<tr>
<th>Diagnoses</th>
<th>Nested cohort—547 patients</th>
<th>37 dropout patients</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>ANA pos (N=288)</td>
<td>ANA neg (N=259)</td>
</tr>
<tr>
<td>Systemic lupus erythematosus</td>
<td>57 19.8</td>
<td>6 2.3</td>
</tr>
<tr>
<td>Inflammatory connective tissue disease</td>
<td>44 15.3</td>
<td>12 4.6</td>
</tr>
<tr>
<td>Inflammatory joint disease</td>
<td>49 17</td>
<td>95 36.7</td>
</tr>
<tr>
<td>Systemic inflammatory disease</td>
<td>10 3.5</td>
<td>17 6.6</td>
</tr>
<tr>
<td>Arthralgia</td>
<td>45 15.6</td>
<td>32 12.35</td>
</tr>
<tr>
<td>Osteoarthritis</td>
<td>25 8.7</td>
<td>29 11.2</td>
</tr>
<tr>
<td>Soft-tissue rheumatism</td>
<td>8 2.8</td>
<td>25 9.65</td>
</tr>
<tr>
<td>Non-rheumatic disease</td>
<td>34 11.8</td>
<td>26 10</td>
</tr>
<tr>
<td>Dermatological disorder</td>
<td>10 3.5</td>
<td>7 2.7</td>
</tr>
<tr>
<td>Unspecified</td>
<td>7 2.4</td>
<td>10 3.9</td>
</tr>
</tbody>
</table>

*Ratio between prevalence of manifestation in ANA positive and ANA negative patients.

ANA and anti-dsDNA antibody assays

The number of patients positive in the different anti-dsDNA assays performed ranged from 33 (6.1%) to 61 (11.1%). Among all 547 sera, a substantial difference between the centres was seen, being the number of CLIFT-positive results 33 (6%), 36 (6.6%) and 45 (8.2%) for Copenhagen, Tromsø and Lund, respectively. A total of 59 patients were positive by at least one CLIFT and 99 by any ELISA assessment, with low agreement of results (k=0.34).

Of the 288 ANA-positive sera, 39 (13.5%) were positive in any CLIFT, while 50 sera (17.4%) were positive in any of the immunoassays (Varelisa, SPADE or EliA). Of the 259 ANA-negative sera, 20 (7.7%) were positive by any CLIFT, and 49 (18.9%) were positive by any immunoassay. One hundred and twenty-four patients (68 ANA positive and 56 ANA negative) were positive at least by one assay, whereof 34 patients (21 ANA positive and 13 ANA negative) were positive by at least one CLIFT and one ELISA assessment.

In general, the CLIFT tests were least often positive. None of the single anti-dsDNA tests was associated with the presence of ANA, with the only exception of the CLIFT performed in Lund. Nonetheless, the positivity for any CLIFT was strongly associated with ANA positivity (table 4). According to our results, having a negative ANA did not rule out having a positive anti-dsDNA test.

Anti-dsDNA antibodies and clinical manifestations

The prevalence of clinical and laboratory manifestations in anti-dsDNA-positive patients are reported in the online supplementary table S3. Malar rash, cutaneous vasculitis, alopecia, leukopenia, lymphopenia, non-haemolytic anaemia, pleuritis, proteinuria and haematuria are more prevalent in patients with combined CLIFT and ELISA positivity (CLIFT+ ELISA+). Sixty-five patients resulted negative by CLIFT and positive by...
ELISA (CLIFT− ELISA+), with higher prevalence of oral or nasal ulcers, tendinitis, xerofthalmia, xerostomia and fibromyalgia. The combination of CLIFT positivity with negative outcome of ELISA (CLIFT+ ELISA−) was more frequent in patients with arthralgia, peripheral arthritis, morning joint stiffness, photosensitivity, Raynaud’s phenomenon, puffy fingers, thrombocytopenia, livedo reticularis, chronic urticaria, discoid LE, asthma, peripheral neuropathy and headache.

Associations between clinical and biochemical manifestations and positivity of anti-dsDNA antibodies are listed in table 5. ORs with 95% CIs are given as crude values (univariate) and as adjusted values (multivariate) for all the individual manifestations significantly associated with presence of anti-dsDNA antibodies, assessed by any CLIFT and any ELISA, namely proteinuria, haematuria, alopecia, leukopenia, thrombocytopenia, cutaneous vasculitis and pleuritis.

Lymphopenia and non-haemolytic anaemia were significantly associated with positive CLIFT, as well as morning stiffness and arthralgia were inversely associated only with a positive ELISA result.

The clinical manifestations most strongly associated with positive result of CLIFT had higher OR in...
ANA-positive patients, except for pleuritis. In patients with any positive ELISA, only OR for cutaneous vasculitis, thrombocytopenia and leukopenia were higher in ANA-positive patients. Having a positive anti-dsDNA test was associated with pleuritis and proteinuria even in ANA-negative patients.

### Table 4
Prevalence of positivity in the anti-dsDNA tests performed

<table>
<thead>
<tr>
<th></th>
<th>All patients</th>
<th>ANA pos (N=288)</th>
<th>ANA neg (N=259)</th>
<th>Rate ratio*</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>n</td>
<td>Per cent n</td>
<td>Per cent n</td>
<td></td>
<td></td>
</tr>
<tr>
<td>CLIFT 1</td>
<td>33</td>
<td>6.1</td>
<td>18</td>
<td>5.8</td>
<td>1.1</td>
</tr>
<tr>
<td>CLIFT 2</td>
<td>36</td>
<td>6.6</td>
<td>22</td>
<td>5.4</td>
<td>1.4</td>
</tr>
<tr>
<td>CLIFT 3</td>
<td>45</td>
<td>8.2</td>
<td>30</td>
<td>5.8</td>
<td>1.8</td>
</tr>
<tr>
<td>Any CLIFT</td>
<td>59</td>
<td>10.8</td>
<td>39</td>
<td>7.7</td>
<td>1.75</td>
</tr>
<tr>
<td>ELIA</td>
<td>61</td>
<td>11.1</td>
<td>27</td>
<td>13.1</td>
<td>0.7</td>
</tr>
<tr>
<td>SPADE</td>
<td>57</td>
<td>10.4</td>
<td>28</td>
<td>11.2</td>
<td>0.9</td>
</tr>
<tr>
<td>Varelisa 1</td>
<td>50</td>
<td>9.1</td>
<td>26</td>
<td>9.3</td>
<td>1</td>
</tr>
<tr>
<td>Varelisa 2</td>
<td>59</td>
<td>10.8</td>
<td>30</td>
<td>11.2</td>
<td>0.9</td>
</tr>
<tr>
<td>Any ELISA</td>
<td>99</td>
<td>18.1</td>
<td>50</td>
<td>18.9</td>
<td>0.9</td>
</tr>
<tr>
<td>Any test</td>
<td>124</td>
<td>22.7</td>
<td>68</td>
<td>21.6</td>
<td>1.1</td>
</tr>
<tr>
<td>CLIFT+ELISA</td>
<td>34</td>
<td>6.2</td>
<td>21</td>
<td>5.0</td>
<td>1.6</td>
</tr>
</tbody>
</table>

Test details are presented in Table 1.

*Ratio between prevalence of positive anti-dsDNA test in ANA positive and ANA-negative patients.

ANA, antinuclear antibodies; CLIFT, Crithidia Lucillia Immunofluorescence Test; SPADE, solution phase anti-dsDNA ELISA.

### Table 5
Association between most relevant clinical manifestations and positive outcome of anti-dsDNA tests (any CLIFT and any ELISA)

<table>
<thead>
<tr>
<th></th>
<th>Any CLIFT positive (39 ANA positive+20 ANA negative)</th>
<th>Any ELISA positive (50 ANA positive+49 ANA negative)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>CRUDE (OR (95% CI))</td>
<td>ADJUSTED (OR (95% CI))</td>
</tr>
<tr>
<td>Peripheral arthritis</td>
<td>1.1 (0.6–2)</td>
<td>0.6 (0.4–1.04)</td>
</tr>
<tr>
<td>Photosensitivity</td>
<td>0.6 (0.2–1.7)</td>
<td>0.5 (0.2–1.2)</td>
</tr>
<tr>
<td>Oral ulcers</td>
<td>0.8 (0.25–2.9)</td>
<td>3.2 (1.3–8)</td>
</tr>
<tr>
<td>Haematuria</td>
<td>3.8 (1.4–10.4)</td>
<td>0.6 (0.1–3.4)</td>
</tr>
<tr>
<td>Proteinuria</td>
<td>14 (5.1–38.4)</td>
<td>13 (2.9–57.7)</td>
</tr>
<tr>
<td>Malar rash</td>
<td>3.1 (0.97–10.2)</td>
<td>1.7 (0.5–5.4)</td>
</tr>
<tr>
<td>Anaemia</td>
<td>3.8 (1.4–10.4)</td>
<td>2.3 (0.5–10.7)</td>
</tr>
<tr>
<td>Leukopenia</td>
<td>6.8 (2.3–20.3)</td>
<td>4.3 (1.1–16)</td>
</tr>
<tr>
<td>Alopecia</td>
<td>4.4 (1.3–15)</td>
<td>4.4 (1.3–15)</td>
</tr>
<tr>
<td>Lymphopenia</td>
<td>10.9 (3.2–37)</td>
<td>2.7 (0.8–9.2)</td>
</tr>
<tr>
<td>Discoid LE</td>
<td>1.2 (0.1–9.8)</td>
<td>0.6 (0.2–1.2)</td>
</tr>
<tr>
<td>Thrombocytopenia</td>
<td>8.8 (2.1–36.2)</td>
<td>3.1 (0.5–20.2)</td>
</tr>
<tr>
<td>Pleuritis</td>
<td>13 (2.1–79.6)</td>
<td>11.1 (1.5–83.8)</td>
</tr>
<tr>
<td>Arthralgia</td>
<td>0.6 (0.3–1.02)</td>
<td>0.5 (0.3–0.8)</td>
</tr>
<tr>
<td>Morning joint stiffness</td>
<td>0.5 (0.2–1.04)</td>
<td>0.4 (0.2–0.7)</td>
</tr>
<tr>
<td>Raynaud’s phenomenon</td>
<td>1.6 (0.8–3.2)</td>
<td>1.2 (0.6–2.2)</td>
</tr>
<tr>
<td>Headache</td>
<td>0.7 (0.3–1.75)</td>
<td>0.8 (0.4–1.6)</td>
</tr>
<tr>
<td>Xerostomia</td>
<td>0.75 (0.3–1.8)</td>
<td>1 (0.5–2)</td>
</tr>
<tr>
<td>Arterial hypertension</td>
<td>0.96 (0.4–2.2)</td>
<td>1 (0.5–1.9)</td>
</tr>
<tr>
<td>Tendinitis</td>
<td>0.2 (0.02–1.2)</td>
<td>1.3 (0.7–2.7)</td>
</tr>
<tr>
<td>Psoriasis</td>
<td>0.6 (0.2–2.2)</td>
<td>0.8 (0.3–1.9)</td>
</tr>
<tr>
<td>Affective disorder</td>
<td>0.7 (0.2–2.2)</td>
<td>0.8 (0.3–2)</td>
</tr>
<tr>
<td>Keratoconjunctivitis sicca</td>
<td>0.4 (0.1–1.8)</td>
<td>0.8 (0.3–9.6)</td>
</tr>
<tr>
<td>Cutaneous vasculitis</td>
<td>6.7 (2.25–20.2)</td>
<td>2.1 (0.5–9.4)</td>
</tr>
<tr>
<td>Asthma or COL</td>
<td>2.4 (1.04–5.5)</td>
<td>1.4 (0.7–3.15)</td>
</tr>
<tr>
<td>Lymphadenopathies</td>
<td>6.5 (1.4–29.7)</td>
<td>1.8 (0.9–3.6)</td>
</tr>
</tbody>
</table>

Crude OR with corresponding 95% CIs in brackets is reported for all the variables. Adjusted OR is reported only for variables significantly associated with positivity of any CLIFT and any ELISA.

ANA, antinuclear antibodies; CLIFT, Crithidia Lucillia Immunofluorescence Test.
The overall association of anti-dsDNA with typical manifestations of SLE

A PCA of the ACR classification criteria5 40 items included in table 4, alopecia (included in the Systemic Lupus International Collaborating Clinics (SLICC) criteria6) and outcomes of anti-dsDNA tests by ELISA (any test positive) and CLIFT (any test positive) is shown as a three-dimensional plot for the three major principal components explaining 16.1%, 10.7% and 8.6%, respectively, of the variance in the dataset. This plot shows how having a positive CLIFT and ELISA(clift+elisa+) is associated with signs of nephropathy, haematological abnormalities and pleuritis; having a single positive test (CLIFT or ELISA) did not cluster with any of the manifestations included in the PC analysis.

The uniqueness of the present study is that the association between clinical phenotypes and presence in serum of anti-dsDNA antibodies has been investigated in consecutive patients with recent onset of rheumatic symptoms, regardless of the diagnosis. By this approach, we aimed to reproduce the usual clinical setting, where the physician, at an early stage, is challenged to formulate a diagnosis and predict the outcome, based on clinical manifestations and suitable diagnostic tools available locally.

This study demonstrates that different techniques for the detection of anti-dsDNA antibodies result in a considerable variation in anti-dsDNA antibody status and relation hereof to various clinical and biochemical manifestations. Shared findings for the regression analyses and the PCA are the relationships between nephropathy and pleuritis with a positive CLIFT. The results obtained for immunoassays were, instead, highly variable and less selective with respect to several typical SLE manifestations. Furthermore, anti-dsDNA antibodies in general were only associated with a few manifestations typically used to classify SLE, and notably with lupus nephritis, where anti-dsDNA antibodies are demonstrated to be involved in the pathogenesis.

These observations raise the discussion about the performance of the various anti-dsDNA detection techniques, the general pathogenic role of the various anti-dsDNA antibodies identified, their clinical associations, and how the syndrome of SLE is currently delineated. Anti-dsDNA antibodies represent a heterogeneous population of antibodies with respect to origin, intrinsic affinity, fine structural DNA specificity and potential to cross-react with non-DNA structures. From this consideration, diverse anti-dsDNA antibody assays may have been designed without implementing this knowledge, but rather with a general view that anti-dsDNA is a homogeneous antibody population. Therefore, in order to analyse this, diverse assay principles are included in this study (table 1). These assays are partly claimed to detect antibodies of low versus high avidity; and those recognised in the CLIFT assays are reported to recognise highly bent structures on DNA, that are disclosed from the general B helical DNA used in some of the ELISA assays. Consistent with this, the kinetoplast DNA has one of the greatest known degrees of stable curvature.41 42 Thus, the assays may disclose antibody binding to DNA structures that are only formed by strong deformations from the more common linear B helical DNA structure. Antibodies recognising the kinetoplast DNA of the haemoflagellate Crithidia luciliae may specifically bind unique structures shared by nucleosomes41 42. This stringent antibody specificity may well reflect structures on eukaryotic nucleosomal DNA that is believed to induce such immune responses in vivo.

We analysed anti-dsDNA antibodies by a variety of anti-dsDNA tests commonly used in clinical practice. CLIFT was performed on all sera at all three laboratories. Each laboratory also performed additional anti-dsDNA antibody assays on all patient sera, depending on their interests and expertise. This approach uncovered a high degree of variation in the reporting of
presence of anti-dsDNA between the individual assays. Even when using a common antigen source (CLIFT) in all laboratories, a high degree of variation in the assay results was demonstrated. It cannot be regarded as acceptable, and it supports the need for further international efforts in assay standardisation, and a broad discussion of what we want to analyse. Do we want to detect as many antibodies as possible, or only those with stringent characteristics, like high avidity and strong correlation with organ manifestations?

The present study shows a high degree of variation in the associations of anti-dsDNA antibodies with clinical and biochemical manifestations, which is in line with previous observations. We do confirm also that immunoassays had higher rates of positivity than CLIFT, as previously reported. Anti-dsDNA antibodies do not represent a well-defined autoantibody entity. Given the multiple and diverse mechanisms for their production, it is less obvious that anti-dsDNA antibodies per se are associated with all aspects of the broad syndrome constituting SLE.

As others and we have observed, individuals may produce anti-dsDNA antibodies without having organ manifestations, like, for example, nephritis or dermatitis. It has been known for decades that not all anti-dsDNA antibodies are pathogenic.

A renal target for potentially nephritogenic anti-dsDNA antibodies has been demonstrated to be extracellular, poorly degraded chromatin fragments in both murine and human lupus nephritis. Antibodies that bind exclusively in ELISA recognise B helical DNA, but not the highly bent structure in the kinetoplast or in the chromatin fragments. Thus, what makes anti-dsDNA antibodies pathogenic is their potential to bind chromatin structures, as reflected in, for example, the CLIFT assay, and not just because they bind any DNA structure. This basic information should encourage us to consider this as a problem, and to develop a new consensus with respect to how we should test for anti-dsDNA antibodies.

In an effort to clinically identify the pathogenic potential of the various anti-dsDNA antibodies measured in this study, we have correlated these findings to clinical and biochemical features of patients referred to our clinics for further evaluation of rheumatic disease. The data support the notion that various anti-dsDNA antibodies impact differently on the classification of SLE.

We focused on whether any of the anti-dsDNA tests were discriminatory towards manifestations included in the current ACR classification criteria. Anti-dsDNA antibodies detected by CLIFT are associated with a clinical picture characterised by proteinuria, haematuria, pleuritis and leukopenia. When the presence of anti-dsDNA antibodies is confirmed by any immunoassay, then the prevalence of malar rash, cutaneous vasculitis, alopecia, lymphopenia and non-haemolytic anaemia seems to increase.

Arthralgia, cutaneous vasculitis, morning stiffness and alopecia are variously associated with the anti-dsDNA tests. Except for alopecia, however, these associations are not strong, and these findings may be spurious. Our results may indicate the existence of a cluster of patients with anti-dsDNA antibodies and diffuse non-scarring alopecia. It supports the recent reinstatement of non-scarring alopecia as a clinical criterion in the SLICC classification criteria of SLE.

With regard to the immunological ACR classification criterion of SLE ‘antibody to native DNA in an abnormal titre’, this study has demonstrated an up to twofold difference in prevalence of positivity, and considerable variation in the clinical association profile between the anti-dsDNA tests performed. This may relate to varying test properties with regard to avidity and structural antibody specificity. Immunoassays are often used to make a preliminary screening of anti-dsDNA antibodies. Positive results are then verified by other more specific assays, such as CLIFT. In our investigation, the positivity of ELISA combined with CLIFT negativity was found in 65 patients, whereof 13 were diagnosed with SLE. A moderately increased prevalence of tendinitis, fibromyalgia, xerothalmia, xerostomia, oral and nasal ulcers was observed, a clinical picture often found in patients affected by Sjögren’s Syndrome. However, these and other parallel findings indicate that currently the anti-dsDNA antibodies cannot be considered precise enough as a classification criterion. Low specificity of ELISA testing for anti-dsDNA antibodies has in newly proposed classification criteria been offset by raising the cut-off to twice above lower laboratory reference range. Whether this is sufficient to generate a valid, clinically relevant anti-dsDNA antibody test situation has not yet been validated. To this end, it is noteworthy that even the CLIFT test, which according to the present results is the anti-dsDNA assay best correlated with a typical nephritic SLE phenotype, had only low diagnostic value in the diagnosis of SLE in this population of patients with recent onset of rheumatic symptoms.

When analysing the ANA-positive subset of the patients, the CLIFT association with clinical phenotypes consisting of proteinuria, haematuria, leukopenia, thrombocytopenia, cutaneous vasculitis and alopecia persisted. On the other hand, pleuritis and proteinuria defined the clinical phenotypes associated with any of the anti-dsDNA tests in ANA-negative patients. These observations suggest that the presence of anti-dsDNA antibodies in ANA-negative patients may be of less clinical significance, supporting the view that testing for antibodies to dsDNA is not indicated in ANA-negative sera. Detection in ANA-negative patients of anti-dsDNA antibodies with low avidity may, in a worst case scenario, lead to incorrect diagnosis and classification.

The purpose of this study was to reach more insight into the linkage between anti-dsDNA antibody detection and clinical and laboratory manifestations without the
restriction posed by existing SLE classification criteria. Our findings provide clinical support for a diagnostic role of CLIFT-determined anti-dsDNA antibodies, however, only in a limited number of key SLE manifestations, proteinuria in particular. These findings challenge the broad role of ELISA-based anti-dsDNA antibody testing in diagnosis and classification of SLE as presently defined.

Author affiliations
1Section of Rheumatology, Department of Clinical Sciences, Lund University, Lund, Sweden
2Department of Biochemistry, Institute of Medical Biology, University of Tromsø, Tromsø, Norway
3Statens Serum Institut, Copenhagen, Denmark
4Department of Rheumatology, Rigshospitalet, Copenhagen University Hospital, Copenhagen, Denmark
5Bone and Joint Research Group, Department of Clinical Medicine, Faculty of Health Science, University of Tromsø, Tromsø, Norway
6Department of Laboratory Medicine, Section of Microbiology, Immunology and Glycobiology, Lund University, Lund, Sweden
7Division of Medicine, Rheumatology Section, Royal Darwin Hospital, Darwin, Northern Territory, Australia

Contributors All the authors contributed to the conception or design of the work, the acquisition, analysis, or interpretation of data for the work. All the authors contributed to drafting the work or revising it critically for important intellectual content. All the authors approved the final version to be published. All the authors achieved agreement for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Funding Supported by academic research funding from the participating centres, the Danish Rheumatism Association (SJ), the Novo Nordisk Research Foundation (SJ), Foundation for Health and Rehabilitation through the Norwegian Rheumatism Organisation (project 2004/20250), Northern Norway Regional Health Authority Medical Research Program (Grants # SFP-100-04, SFP-101-04), University of Tromsø as a Milieu support (OPR), Swedish Rheumatic Patients’ Association (AAB), The Foundation of King Gustav V 80-year fund (AAB).

Competing interests None.

Ethics approval Tromsø, Norway: Regional Committees for Medical and Health Research Ethics - REC North, project no. P. Rek Nord 03/2004; Lund, Sweden: Research Ethics Committee, Medical Faculty, Lund University, project no. LU 30-03 (LU-P12-03); Copenhagen, Denmark: Research Ethics Committees for Copenhagen and Frederiksberg, project no. (KF) 01-024/03.

Provenance and peer review Not commissioned; externally peer reviewed.

Data sharing statement No additional data are available.

Open Access This is an Open Access article distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 3.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the Creative Commons Attribution Non Commercial (CC BY-NC 3.0) license, and the work is properly cited and the use is non-commercial. See: http://creativecommons.org/licenses/by-nc/3.0/.

REFERENCES


41. Werle E, Blazek M, Fiehn W. The clinical significance of measuring different anti-dsDNA antibodies by using the Farr assay, an enzyme immunosassay and a Crithidia luciliae immunofluorescence test.


### General symptoms
- Noninfectious fever
- Weight loss
- Anorexia
- Other

### Vascular system
- Raynauds phenomenon
- Venous thrombosis
- Arterial thrombosis
- Cerebral infarction
- Transient ischemic attack
- Avascular bone necrosis
- Claudicatio intermittens
- Arterial hypertension
- >1 spontaneous abortion
- Other

### Hematology
- Non-hemolytic anemia
- Immune hemolytic anemia
- Leucocytopenia
- Lymphocytopenia
- Thrombocytopenia
- Lymphadenopathy
- Other

### General symptoms
- Other

### Other

### Skin
- Butterfly rash
- Photosensitivity
- Discoid LE
- Subacute LE
- Alopecia
- Purpura
- Cutaneous vasculitis
- Genital ulcers
- Chronic urticaria
- Telangiectasias
- Rheumatoid nodules
- Panniculitis
- Bullae
- Periorbital edema/cyanosis
- Gottron’s sign
- Livedo reticularis
- Pitting scars
- Proximal scleroderma
- Psoriasis
- Other

### Joints and muscles
- Morning stiffness
- Palindromic arthritis
- Puffy fingers
- Peripheral arthritis
- Axial arthritis
- Arthralgia
- Erosions on x-ray
- Tendinitis
- Myositis
- Fibromyalgia
- Other

### Eyes
- Keratoconjunctivitis sicca
- Scleritis/episcleritis
- Anterior uveitis
- Posterior uveitis
- Retinal vasculitis
- Other

### Vascular system
- Other

### Other

### Joint and muscles
- Other

### Eyes
- Other

### Other

### Laboratory
- ESR (mm/h)
- CRP (mg/L)
- Hgb (mmol/L)
- Creatinin (µmol/L)

---

**Scandinavian anti-DNA Study (ScantiDNA)**

**Clinical case record form**

**Patient no.:**

**Date of birth:** __/__/__

**Date of inclusion:** __/__/__

**Smoking:** yes / no / previous

**Gender:** M / F

**Race:** White / Asian / African / Hispanic
- Scandinavian anti-DNA Study
- Non-infectious fever
- Weight loss
- Anorexia
- Other

**Other:**

**Height:** cm  **Weight:** kg

**Manifestation:** absent = 0; present = X  inactive = I; unknown = U  Onset of manifestation: __/__/__

**Noninfectious fever**
- Yes
- No
- Previous

**Weight loss**
- Yes
- No
- Previous

**Anorexia**
- Yes
- No
- Previous

**Other**
- Yes
- No
- Previous

**Skin**

- Butterfly rash
- Photosensitivity
- Discoid LE
- Subacute LE
- Alopecia
- Purpura
- Cutaneous vasculitis
- Genital ulcers
- Chronic urticaria
- Telangiectasias
- Rheumatoid nodules
- Panniculitis
- Bullae
- Periorbital edema/cyanosis
- Gottron’s sign
- Livedo reticularis
- Pitting scars
- Proximal scleroderma
- Psoriasis
- Other

**Heart**
- Pericarditis
- Myocarditis
- Endocarditis
- Angina
- Myocardial infarction
- Arrhythmia
- Cardiac failure
- Other

**Pulmonary**
- Pleuritis
- Alveolitis/fibrosis
- Pulmonary hypertension
- Asthma bronch/COLD
- Other

**Kidney**
- Proteinuria
- Hematuria
- Sterile pyuria
- Cellular casts
- Glomerulonephritis on biopsy
- WHO class
- Other

**Gastrointestinal**
- Oral/nasal ulcers
- Xerostomia
- Sterile peritonitis/ascites
- Intestinal vasculitis
- Autoimmune hepatitis
- Pruritus biliary cirrhosis
- Celiac disease
- Other

**Hematology**
- Non-hemolytic anemia
- Immune hemolytic anemia
- Leucocytopenia
- Lymphocytopenia
- Thrombocytopenia
- Lymphadenopathy
- Other

**Neuropsychiatry**
- Headache
- Cognitive dysfunction
- Seizures/cholesterosis
- Peripheral neuropathy
- Cranial nerve affection
- Transverse myelopathy
- Organic brain syndrome
- Psychosis
- Affective disorder
- Other

**Endocrinology**
- Thyroiditis
- Type 1 diabetes mellitus
- Amenorrhea
- Other

**Infections**
- >2 major infections

**Malignancy**
- Previous malignancy
- Type
- Current malignancy
- Type

**Medication last yr**
- Heart
- Blood pressure
- Epilepsia
- Others

**Laboratory**
- ESR (mm/h)
- CRP (mg/L)
- Hgb (mmol/L)
- Creatinin (µmol/L)
<table>
<thead>
<tr>
<th>Term</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Non-infectious fever</td>
<td>The objective determination of an elevation of body temperature above the normal range (i.e. 37 ± 1 °C).</td>
</tr>
<tr>
<td>Weight loss</td>
<td>Decrease in body weight that is not voluntary.</td>
</tr>
<tr>
<td>Anorexia</td>
<td>Reduction or loss of appetite or desire for food.</td>
</tr>
<tr>
<td>Butterfly (Malar) Rash</td>
<td>Diffuse or patchy erythema of the malar eminence(s). Lesions may be flat or raised, involving cheeks and/or the bridge of the nose but tending to spare the nasolabial folds; may be unilateral and may involve adjacent or other areas.</td>
</tr>
<tr>
<td>Photosensitivity</td>
<td>An unusual skin reaction from exposure to sunlight (typically UV-B). Examples would be persistent erythema, edema, urticaria or vesicular-bullous lesions, located in sun-exposed areas.</td>
</tr>
<tr>
<td>Discoid LE (Discoid rash)</td>
<td>Rash occurring predominantly (but not exclusively) in sun exposed areas and characterized by erythematous, raised patches with adherent keratotic scaling and follicular plugging; atrophic scarring, telangiectasias, hyperpigmentation (peripheral) and hypopigmentation (central) may be present in older lesions.</td>
</tr>
<tr>
<td>Subacute LE</td>
<td>Widespread photosensitive, nonscarring eruptions, either papulosquamos (psoriasiform) or annular.</td>
</tr>
<tr>
<td>Alopecia</td>
<td>An abnormal patchy or diffuse loss of hair, particularly scalp hair, non-scarring.</td>
</tr>
<tr>
<td>Purpura</td>
<td>Intracutaneous or subcutaneous hemorrhage as evidenced by red to dark purple areas in the skin.</td>
</tr>
<tr>
<td>Cutaneous vasculitis</td>
<td>Confirmed by skin biopsy or convincing clinical presentation when present on acral sites where biopsy is not feasible.</td>
</tr>
<tr>
<td>Genital ulcers</td>
<td>A break in the skin or mucous membrane found on the penis, scrotum, labia, vestibule or vagina. Lesions may be painful or painless, single or multiple, recurrent or persistent.</td>
</tr>
<tr>
<td>Chronic urticaria</td>
<td>A disorder of the superficial skin consisting of well circumscribed discrete wheals with erythematous raised serpiginous borders and blanched centers. It is usually intensely pruritic, and may be localized or generalized.</td>
</tr>
<tr>
<td>Teleangiectasias</td>
<td>Visible macular dilatation of superficial cutaneous blood vessels. These blood vessels collaps upon pressure and fill slowly when pressure is released.</td>
</tr>
<tr>
<td>Rheumatoid nodules</td>
<td>Firm, usually painless lumps of variable size found in patients with rheumatoid arthritis. Rheumatoid nodules are commonly found over areas subject to mechanical trauma (e.g., elbows, heels, walls of olecranon bursa), and occasionally in various internal organs such as lungs and heart.</td>
</tr>
<tr>
<td>Panniculitis</td>
<td>Nodular, subcutaneous angitis with fat-cell necrosis or clinically erythema nodosum.</td>
</tr>
</tbody>
</table>
Bullae  Vesicular elevation of the cuticle containing transparent watery fluid.

Periorbital oedema or cyanosis  Violaceous periorbital erythema often with upper eye lid swelling or periorbital oedema.

Gottron’s sign  Erythematous patches. Scaly hyperemic patches present over the extensor surface of the knuckles (DIP, PIP and MCP). The eruption may have atrophic features as well.

Livedo reticularis  Reddish/cyanotic reticular discoloration of the skin. Appears on legs, arms, adn torso.

Pitting scars  Digital scarring with loss of substance after acral ulcers.

Proximal scleroderma  Thickening, tightening, nonpitting induration of the skin of both extremities proximal to the MCP (or MTP) joints and the trunk (anterior chest, abdomen, upper or lower back or flanks).

Psoriasis  A chronic hyperkeratotic recurrent skin disorder most often characterized by somewhat raised, sharply margined papules or plaques which are scaling and distributed predominantly on the scalp, elbows, knees, chest, umbilicus, back and buttocks. Frequent involvement of the fingernails and toenails is present.

Morning stiffness  The subjective complaint of localized or generalized lack of easy mobility of the joints upon arising.

Puffy fingers  A diffuse, usually nonpitting increase in soft tissue mass of the digits extending beyond the normal confines of the joint capsule.

Polyarthritis  Symmetric involvement of more than three joints with clinical signs of synovitis.

Oligoarthritis  Clinical signs of synovitis in three or less joints, often asymmetric.

Axial arthritis  Radiographic signs of sacroiliitis or inflammatory spondylarthropathy.

Arthralgia  Subjective reporting of pain in the joints.

Erosions on x-ray  An erosion is a localized area of bone destruction at or near the joint surface.

Tendinitis  Tenosynovitis determined clinically or by ultrasound.

Fibromyalgia  Widespread pain and tender points as defined by ACR.

Myositis  Muscle weakness accompanied by elevated plasma levels of muscle enzymes. The myopathy is further confirmed by muscle biopsy and/or electromyography.

Keratoconjunctivitis sicca  Confirmed by ophthalmological evaluation using Schirmer test, break up time or Rose Bengal dye.

Scleritis or Episcleritis  Clinical signs of inflammation of the sclera and episclera.

Anterior uveitis  Inflammation of the iris (iritis) or of the iris and the ciliary body (iritocyclitis) is referred to as anterior uveitis and results in photophobia, some decrease in visual acuity, and a variable degree of ocular pain. In contrast to the acute anterioir uveitis, the chronic anterior uveitis associated with juvenile rheumatoid arthritis is frequently asymptomatic.
Posterior uveitis
Inflammation of the choroid usually involves the retina and the term posterior uveitis and chorioretinitis are often used interchangeably.

Retinal vasculitis
Observed by ophthalmoscopy or by a fluorescein angiogram.

Raynaud’s phenomenon
Sudden, reversible "dead white" pallor of an acral structure (e.g., fingers, whole hand, toes, tip of nose, earlobe or tongue), precipitated by cold exposure or emotion.

Venous or arterial thrombosis
Thrombosis should only be recognized when clinical suspicion is confirmed by relevant paraclinical method.

Cerebral infarction
Confirmed by relevant neuroimaging technique (CT, MRI)

Transient ischemic attack
Clinical picture of cerebral infarction with full remission of symptoms within 24 hours.

Avascular bone necrosis
Confirmed by conventional radiography, CT or MRI.

Claudicatio intermittens
Muscle pain (ache, cramp, numbness or sense of fatigue), classically in the calf muscle, which occurs during exercise and is relieved by a short period of rest.

Arterial hypertension
Blood pressure > 140/90 and/or commencement of antihypertensive treatment.

>1 spontaneous abortion
Note time for 2nd spontaneous abortion.

Endocarditis
Non-infectious endocarditis verified by ultrasonography.

Myocarditis
Myocarditis may cause arrhythmias and/or cardiac failure and confirmed by myocardial biopsy.

Pericarditis
Pericardial pain with at least 1 of the following: rub, effusion, or electrocardiogram or echocardiogram confirmation.

Angina pectoris
Severe chest pain due cardiac ischemia without signs of myocardial infarction.

Myocardial infarction
Confirmed by elevated cardiac enzyme levels and electrocardiogram.

Arrhythmia
Atrial or ventricular arrhythmias, conduction disturbances documented by electrocardiogram.

Cardiac failure
Includes both right and left ventricular failure.

Pleuritis
Pleuritic chest pain with pleural rub or effusion, or pleural thickening.

Alveolitis / fibrosis
Active inflammatory alveolitis and/or pulmonary fibrosis confirmed by bronchoalveolar lavage, high resolution CT or conventional radiography.

Pulmonary hypertension
Mean pulmonary artery pressure exceeding 15 mmHg calculated by means of echocardiography or measured by cardiac catheterization.

Asthma or COLD
Confirmed by spirometry indicating intermittent or chronic obstructive ventilatory pattern, FEV1/FVC<70% of expected.

Proteinuria
>0.5 gram/24hours.

Hematuria
>5 red blood cells/high power field.

Sterile pyuria
>5 white blood cells/high power field.

Cellular casts
Heme-granular or red blood cell casts.

Glomerulonephritis
verified by renal biopsy.

WHO-class
1: normal/minimal changes
2: mesangioproliferative GN
3: focal, segmental proliferative GN
4: diffuse proliferative GN
5: membraneous GN
6: end-stage GN

Oral or nasal ulcers
Erosions, superficial or deep, of the buccal, labial, lingual, palatal, pharyngeal, or nasal mucosa. They may be painful or painless.

Xerostomia
Oral dryness based on salivary gland destruction documented by sialometry, salivary scintigraphy or salivary gland biopsy.

Sterile peritonitis or ascites
Documented by imaging or puncture.

Intestinal vasculitis
Confirmed by abdominal angiography or histologically.

Autoimmune hepatitis
Exclusion of viral etiology and confirmed by liver biopsy.

Primary biliary cirrhosis
Elevated serum levels of alkaline phosphatase, often anti-mitochondrial antibodies. Confirmed by liver biopsy.

Celiac disease
1) evidence of malabsorption, 2) abnormal jejunal biopsy showing characteristic changes of the villi, and 3) clinical, and serological improvement after institution of a gluten-free diet.

Non-hemolytic anemia
Blood level of hemoglobin below lower normal range without signs of hemolysis. Evaluation of hemolysis may include reticulocyte count, serum levels of LDH, free hemoglobin and haptoglobin.

Immunohemolytic anemia
Blood level of hemoglobin below lower normal range and positive direct antiglobulin (Coombs') test for autoantibodies directed against the rbc membrane antigens.

Leucocytopenia
<3.000 white blood cells x 10^9 / L.

Lymphocytopenia
Below local lower normal range.

Thrombocytopenia
<100.000 platelets x 10^9 / L.

Lymphadenopathy
An enlargement of lymph nodes greater than normal for the particular region examined.

Headache
Includes: Migraine, Tension headache, Cluster headaches, Pseudotumor cerebri (benign intracranial hypertension) and Intractable non-specific headache. Lupus headache: Severe persistent headache; may be migrainous, but must be non-responsive to narcotic analgesia.

Cognitive dysfunction
The types of cognitiv deficits patients manifest include complex attention, aspects of memory (e.g. learning and recall), visual-spatial processing, language (e.g. verbal), psychomotor speed. Cognitive dysfunction can range from mild impairment to severe dementia. It represents a decline from a previously higher level of functioning and may impede social, educational or occupational functioning. Subjective complaints of cognitive dysfunction are common, although not always objectively verifiable.
Neuropsychological testing is the diagnostic procedure of choice for suspected cognitive dysfunction.

**Aseptic meningitis**
A clinical syndrome of fever, headache and meningeal irritation with CSF pleocytosis and negative cultures.

**Seizures or chorea**
Seizures: Abnormal paroxysmal neuronal discharge in the brain causing abnormal function. Seizures may occur with or without the loss of consciousness. Seizures are divided in two groups, partial and generalized. Partial seizures have clinical and electroencephalographic evidence of a focal onset: the abnormal discharge usually arises in a portion of a hemisphere and may spread to the rest of the brain during a seizure. Primary generalized seizures have no interictal evidence on EEG of focal onset. A generalized seizure can be primary or secondary.

**Chorea**
Chorea consists of irregular, involuntary and jerky movements, that may involve any portion of the body in random sequence. Each movement is brief and unpredictable.

**Peripheral neuropathy**
Mononeuropathy single/multiplex: Disturbance of the function of one or more peripheral nerve(s). Weakness and paralysis can be due to either conduction block in the motor nerve fibers or to axonal loss. Conduction block is related to demyelinisation with preservation of axon continuity. Remyelinisation can be rapid and complete. If axonal interruption takes place, axonal degeneration occurs below the site of interruption and recovery is often slow and incomplete. Sensory symptoms and sensory loss may affect all modalities or be restricted to certain forms of sensation.

**Plexopathy**
A disorder of the brachial or lumbosacral plexus producing muscle weakness, sensory deficit and/or reflex change that do not correspond to the territory of a single root or nerve.

**Polyneuropathy**
Acute or chronic disorder of sensory and motor peripheral nerves with variable tempo characterized by symmetry of symptoms and physical findings in a distal distribution.

**Autonomic neuropathy**
A disorder of the autonomous nervous system which gives rise to orthostatic hypotension, sphincteric erectile/ejaculatory dysfunction, anhidrosis, heat intolerance, constipation.

**Cranial nerve affection**
A clinical syndrome affecting the specific sensory and/or motor function of the cranial nerve(s).

**Transverse myelopathy**
Disorder of the spinal cord characterized by rapidly evolving paraparesis and/or sensory loss, with a demonstrable motor and/or sensory cord level and/or sphincter involvement.

**Organic brain syndrome**
Altered mental function with impaired orientation, memory or other intellectual function, with rapid onset and fluctuating clinical features, inability to sustain attention to environment, plus at least two days of the following:
perceptual disturbance, incoherent speech, insomnia or daytime drowsiness, or increased or decreased psychomotor activity.

Psychosis
Severe disturbance in the perception of reality characterized by delusions and/or hallucinations.

Affective disorder
Prominent or persistent disturbance in mood characterized by either: - Depressed mood or markedly diminished interest or pleasure in almost all activities or – Elevated, expansive or irritable mood.

Thyreoditis
Diagnosis supported by thyroid scintigraphy, thyroid hormone status depends on phase of thyreoiditis.

Type 1 diabetes mellitus
Immune-mediated diabetes mellitus characterized by young age of onset, normal to wasted body habitus, low to absent plasma insulin, and high suppressible plasma glucagon.

Amenorrea
Failure of menarche by age 16 or absence of menstruation for 6 months in a woman with previous periodic menses.

>2 major infections
Note time of 3rd infection requiring hospitalization.
Table S3. Number of anti-dsDNA positive patients and prevalence of their clinical manifestations.

<table>
<thead>
<tr>
<th>Assays</th>
<th>Any CLIFT +</th>
<th>Any ELISA +</th>
<th>CLIFT + ELISA –</th>
<th>CLIFT – ELISA +</th>
<th>CLIFT+ ELISA+</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total number of patients</td>
<td>59</td>
<td>99</td>
<td>25</td>
<td>65</td>
<td>34</td>
</tr>
</tbody>
</table>

Manifestations among SLE classification criteria:

<table>
<thead>
<tr>
<th>Manifestation</th>
<th>Any CLIFT +</th>
<th>Any ELISA +</th>
<th>CLIFT + ELISA –</th>
<th>CLIFT – ELISA +</th>
<th>CLIFT+ ELISA+</th>
</tr>
</thead>
<tbody>
<tr>
<td>Peripheral arthritis (%)</td>
<td>30.5</td>
<td>21.2</td>
<td>44</td>
<td>21.5</td>
<td>20.6</td>
</tr>
<tr>
<td>Photosensitivity (%)</td>
<td>6.8</td>
<td>6.1</td>
<td>8</td>
<td>6.1</td>
<td>5.9</td>
</tr>
<tr>
<td>Oral/nasal ulcers (%)</td>
<td>5.1</td>
<td>9.1</td>
<td>4</td>
<td>10.8</td>
<td>5.9</td>
</tr>
<tr>
<td>Haematuria (%)</td>
<td>10.2</td>
<td>8.1</td>
<td>4</td>
<td>4.6</td>
<td>14.7</td>
</tr>
<tr>
<td>Proteinuria (%)</td>
<td>16.9</td>
<td>13.1</td>
<td>4</td>
<td>6.1</td>
<td>26.5</td>
</tr>
<tr>
<td>Malar rash (%)</td>
<td>6.8</td>
<td>4</td>
<td>0</td>
<td>0</td>
<td>11.8</td>
</tr>
<tr>
<td>Leukopenia (%)</td>
<td>10.2</td>
<td>8.1</td>
<td>8</td>
<td>6.1</td>
<td>11.8</td>
</tr>
<tr>
<td>Alopecia (%)</td>
<td>6.8</td>
<td>5</td>
<td>4</td>
<td>3.1</td>
<td>8.8</td>
</tr>
<tr>
<td>Lymphopenia (%)</td>
<td>10.2</td>
<td>4</td>
<td>8</td>
<td>0</td>
<td>11.8</td>
</tr>
<tr>
<td>Discoid LE (%)</td>
<td>1.7</td>
<td>1</td>
<td>4</td>
<td>1.5</td>
<td>0</td>
</tr>
<tr>
<td>Thrombocytopenia (%)</td>
<td>6.8</td>
<td>4</td>
<td>8</td>
<td>3.1</td>
<td>5.9</td>
</tr>
<tr>
<td>Pleuritis (%)</td>
<td>5.1</td>
<td>4</td>
<td>0</td>
<td>1.5</td>
<td>8.8</td>
</tr>
</tbody>
</table>

Other manifestations:

<table>
<thead>
<tr>
<th>Manifestation</th>
<th>Any CLIFT +</th>
<th>Any ELISA +</th>
<th>CLIFT + ELISA –</th>
<th>CLIFT – ELISA +</th>
<th>CLIFT+ ELISA+</th>
</tr>
</thead>
<tbody>
<tr>
<td>Arthralgia (%)</td>
<td>44.1</td>
<td>42.4</td>
<td>60</td>
<td>47.7</td>
<td>32.3</td>
</tr>
<tr>
<td>Morning joint stiffness (%)</td>
<td>13.6</td>
<td>11.1</td>
<td>24</td>
<td>13.8</td>
<td>5.9</td>
</tr>
<tr>
<td>Raynaud’s phenomenon (%)</td>
<td>18.6</td>
<td>15.1</td>
<td>20</td>
<td>13.8</td>
<td>17.6</td>
</tr>
<tr>
<td>Headache (%)</td>
<td>10.2</td>
<td>11.1</td>
<td>20</td>
<td>15.4</td>
<td>2.9</td>
</tr>
<tr>
<td>Xerostomia (%)</td>
<td>10.2</td>
<td>13.1</td>
<td>12</td>
<td>15.4</td>
<td>8.8</td>
</tr>
<tr>
<td>Arterial hypertension (%)</td>
<td>11.9</td>
<td>12.1</td>
<td>12</td>
<td>12.3</td>
<td>11.8</td>
</tr>
<tr>
<td>Condition</td>
<td>Value 1</td>
<td>Value 2</td>
<td>Value 3</td>
<td>Value 4</td>
<td>Value 5</td>
</tr>
<tr>
<td>-----------------------------------------------</td>
<td>---------</td>
<td>---------</td>
<td>---------</td>
<td>---------</td>
<td>---------</td>
</tr>
<tr>
<td>Tendinitis (%)</td>
<td>1.7</td>
<td>11.1</td>
<td>0</td>
<td>15.4</td>
<td>2.9</td>
</tr>
<tr>
<td>Psoriasis (%)</td>
<td>5.1</td>
<td>4</td>
<td>8</td>
<td>4.6</td>
<td>2.9</td>
</tr>
<tr>
<td>Affective disorder (%)</td>
<td>5.1</td>
<td>6.1</td>
<td>8</td>
<td>7.7</td>
<td>2.9</td>
</tr>
<tr>
<td>Keratoconjunctivitis sicca (%)</td>
<td>3.4</td>
<td>6.1</td>
<td>4</td>
<td>7.7</td>
<td>2.9</td>
</tr>
<tr>
<td>Asthma bronchiale (%)</td>
<td>13.6</td>
<td>9.1</td>
<td>16</td>
<td>7.7</td>
<td>11.8</td>
</tr>
<tr>
<td>Puffy fingers (%)</td>
<td>6.8</td>
<td>6.1</td>
<td>12</td>
<td>7.7</td>
<td>2.9</td>
</tr>
<tr>
<td>Thyreoiditis (%)</td>
<td>5.1</td>
<td>5</td>
<td>4</td>
<td>4.6</td>
<td>5.9</td>
</tr>
<tr>
<td>Peripheral neuropathy (%)</td>
<td>5.1</td>
<td>4</td>
<td>8</td>
<td>4.6</td>
<td>2.9</td>
</tr>
<tr>
<td>Weight loss (%)</td>
<td>6.8</td>
<td>4</td>
<td>8</td>
<td>3.1</td>
<td>5.9</td>
</tr>
<tr>
<td>Axial arthritis (%)</td>
<td>5.1</td>
<td>4</td>
<td>4</td>
<td>3.1</td>
<td>5.9</td>
</tr>
<tr>
<td>Fibromyalgia (%)</td>
<td>1.7</td>
<td>5</td>
<td>0</td>
<td>6.1</td>
<td>2.9</td>
</tr>
<tr>
<td>Anaemia, non-haemolytic (%)</td>
<td>10.2</td>
<td>4</td>
<td>8</td>
<td>0</td>
<td>11.8</td>
</tr>
<tr>
<td>Non-infectious fever (%)</td>
<td>1.7</td>
<td>2</td>
<td>4</td>
<td>3.1</td>
<td>0</td>
</tr>
<tr>
<td>Arrhythmia (%)</td>
<td>3.4</td>
<td>4</td>
<td>0</td>
<td>3.1</td>
<td>5.9</td>
</tr>
<tr>
<td>Cutaneous vasculitis (%)</td>
<td>10.2</td>
<td>8.1</td>
<td>0</td>
<td>3.1</td>
<td>17.6</td>
</tr>
<tr>
<td>Miscarriage/abortion (%)</td>
<td>3.4</td>
<td>1</td>
<td>4</td>
<td>0</td>
<td>2.9</td>
</tr>
<tr>
<td>Chronic urticaria (%)</td>
<td>1.7</td>
<td>0</td>
<td>4</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Livedo reticularis (%)</td>
<td>3.4</td>
<td>0</td>
<td>8</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Palindromic arthritis (%)</td>
<td>3.4</td>
<td>2</td>
<td>4</td>
<td>1.5</td>
<td>2.9</td>
</tr>
</tbody>
</table>